

# Translational PK/PD Modelling

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Data & Clarity

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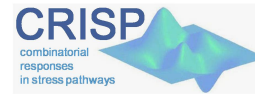


# About me



Mathematical and statistical modelling, Bioinformatics

- Chemical Engineering, BE, 1998-2002
- Biological and Chemical Engineering, MSc, 2002-2003
- Cancer Bioinformatics, 2003-2005
- Systems Biology, PhD, 2005-2009
- Systems Biology, Postdoc, 2009-2011
- Physiological Modeller & PK/PD Modeller, 2011-2015
- PK/PD Modelling Lab Head, 2016-2018
- PK/PD Modeller & Data Scientist, Aug 2018 - now



# Aims

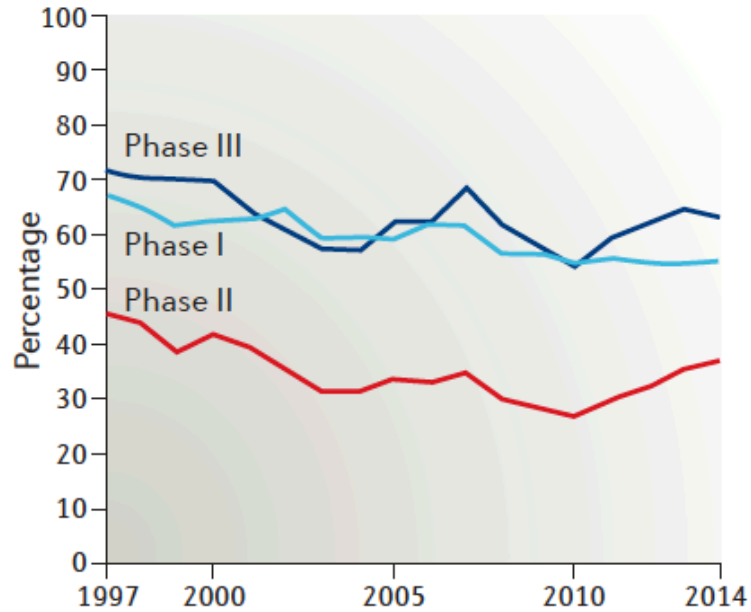
- **Review** the pain points in drug R&D
- **Evaluate** the growth of *in vivo* tumour models
- **Translational** PK/PD modelling: Value proposition, examples
- **Gene signature** for tumour growth

# 1. THE REASON WHY?

# Probability of Success: Phase-by-phase

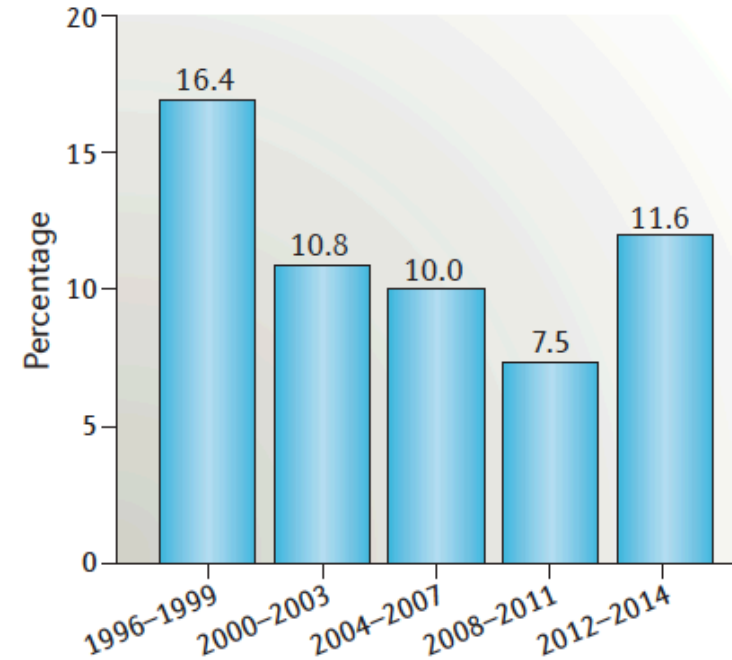
**a Success rates by phase**

Percentage likelihood of moving to next phase, 3-year rolling average\*



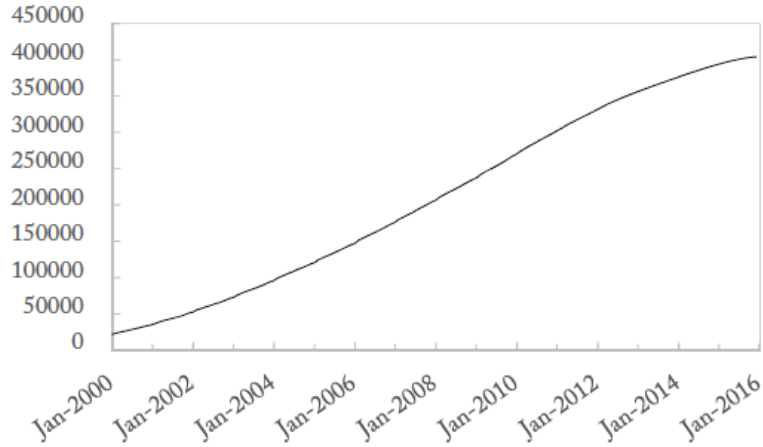
**b Cumulative success rate Phase I to launch**

Percentage likelihood of moving from Phase I to launch



Smietana *et al.* (2016) Trends in clinical success rates. *Nat Rev Drug Disc.* 15: 379-380

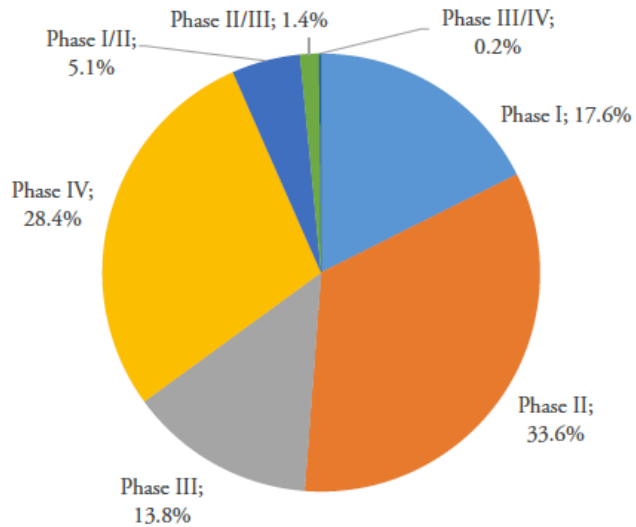
# Clinical trials 2000-2015



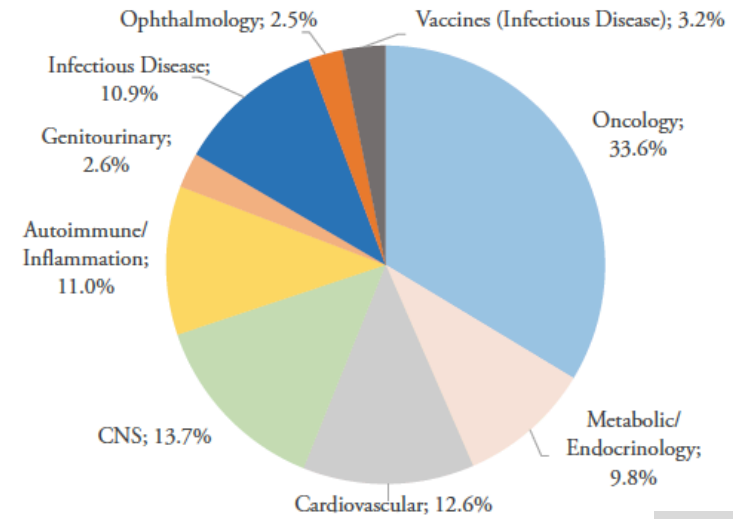
(a) Cumulative number of trials over time



(b) Increase in the number of trials over time



(c) Proportion of trials by phases

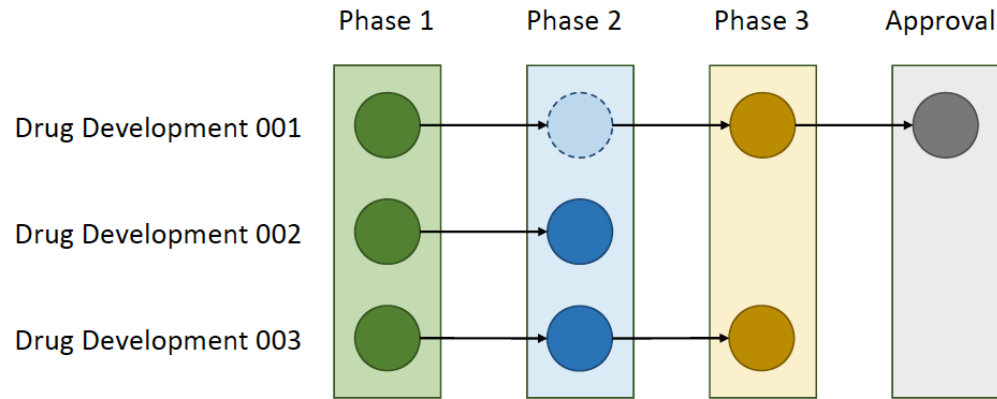


(d) Proportion of trials by therapeutic groups

~186,000 trials in total

Lo *et al.* (2018) Estimation of clinical trial success rates and related parameters. *Biostatistics*. p 1–14

# Probability of Success (POS)

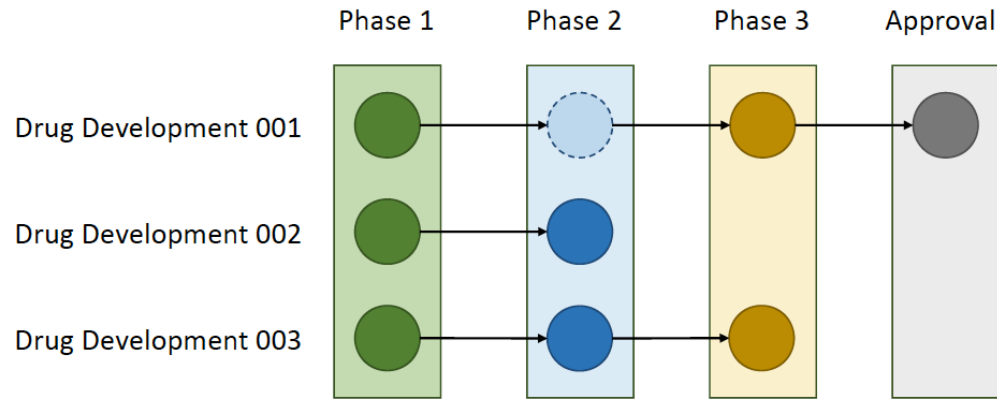


## • Phase-by-phase counting

- $POS_{1,2} = 1$
- $POS_{2,3} = \frac{1}{2}$
- $POS_{3,App} = \frac{1}{2}$
- $POS_{1,App} = 1 \times \frac{1}{2} \times \frac{1}{2} = \frac{1}{4}$
- Widely used in the past
- Ignore missing trials

Lo *et al.* (2018) Estimation of clinical trial success rates and related parameters. *Biostatistics*. p 1–14

# Probability of Success (POS)



## • Path-by-path counting

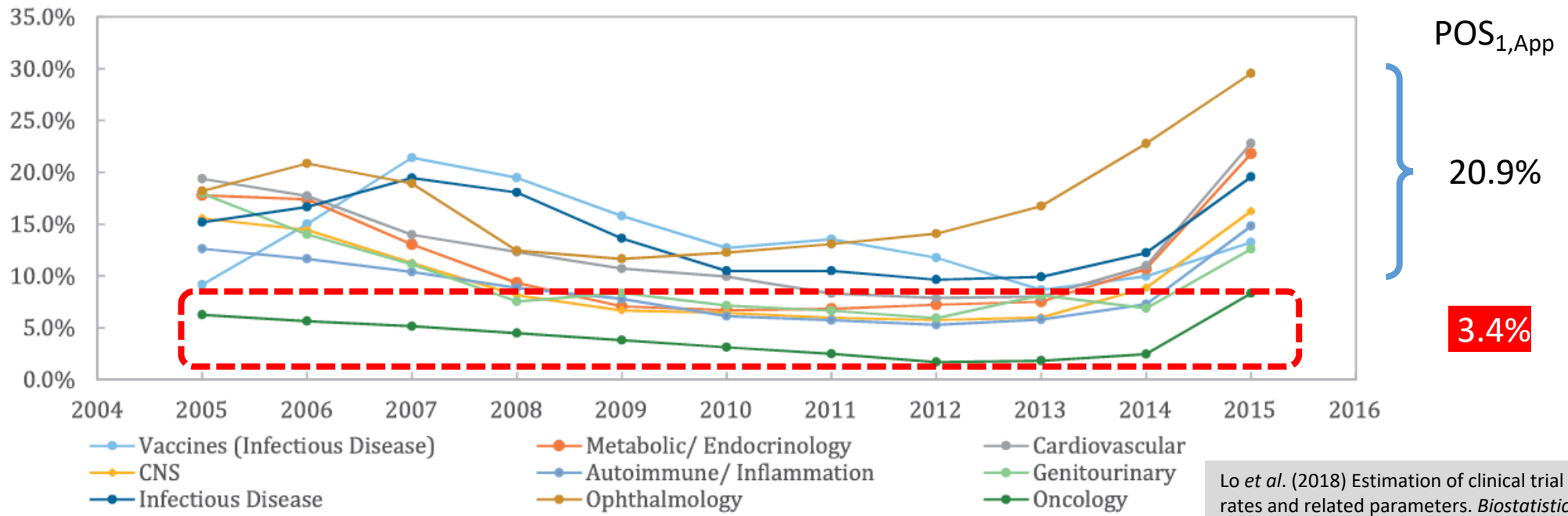
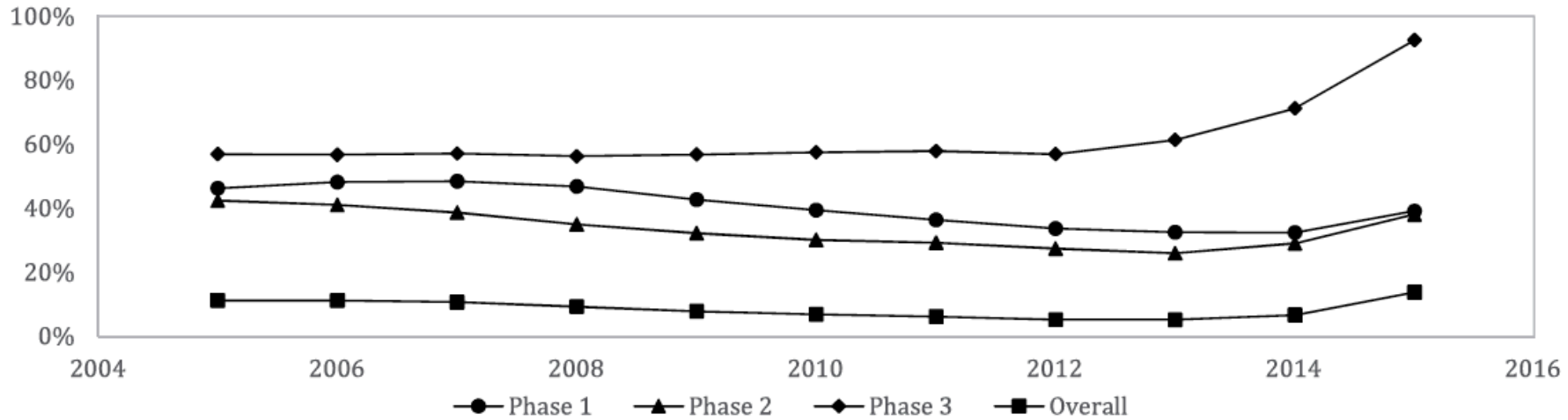
- Missing Phase 2 is inferred
- $POS_{1,2} = 1$
- $POS_{2,3} = \frac{2}{3}$
- $POS_{3,App} = \frac{1}{2}$
- $POS_{1,App} = 1 \times \frac{2}{3} \times \frac{1}{2} = \frac{1}{3}$
- Used by Lo *et al*
- Considers missing, in progress and terminated trials
- More accurate description than phase-by-phase

Lo *et al.* (2018) Estimation of clinical trial success rates and related parameters. *Biostatistics*. p 1–14





# Cancer drug programmes are more risky (on average) than previously thought



Lo et al. (2018) Estimation of clinical trial success rates and related parameters. *Biostatistics*. p 1-14



# Cancer drug: Phase 2 is key



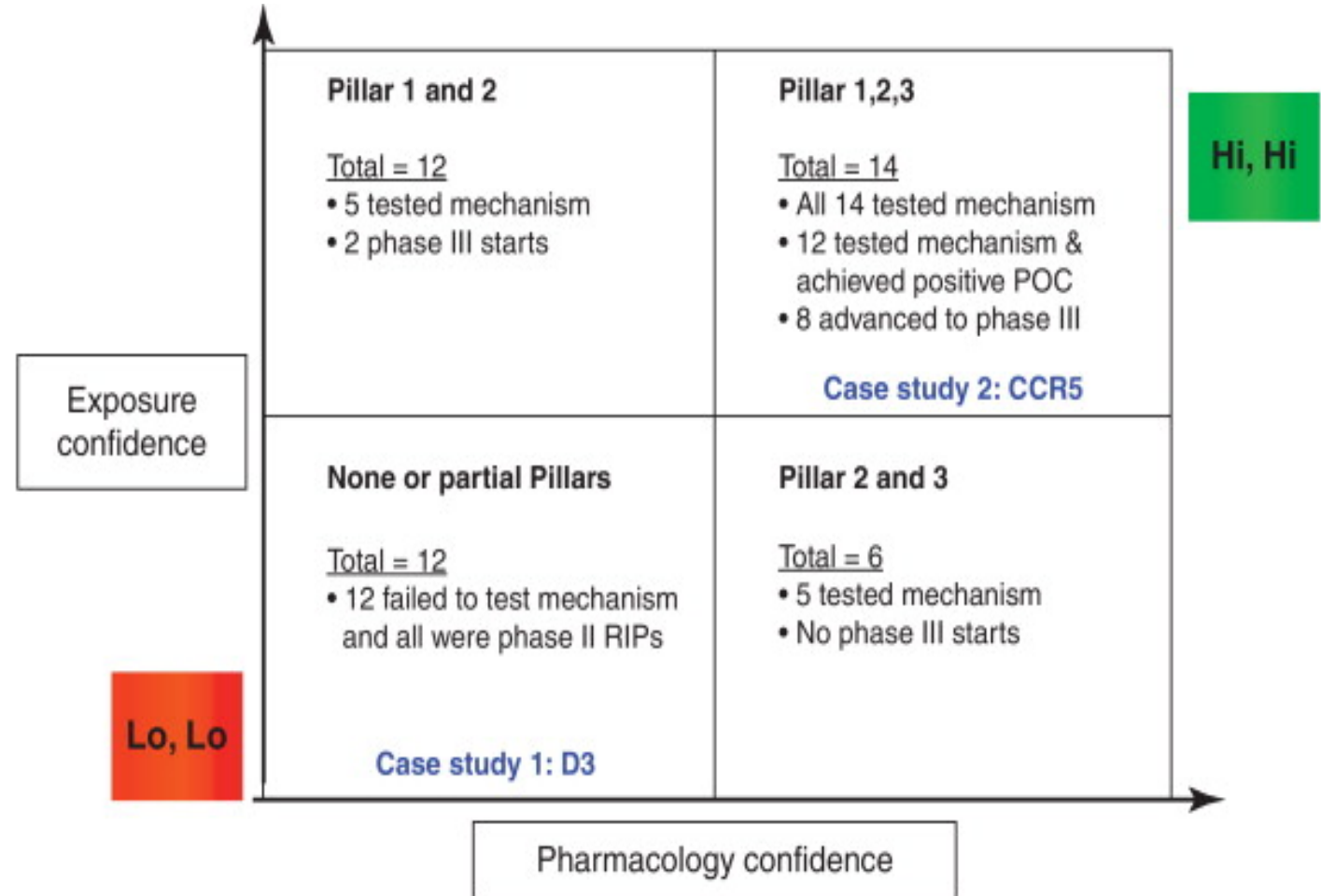
All indications (industry)

Therapeutic group	Phase 1 to Phase 2		Phase 2 to Phase 3			Phase 3 to Approval		Overall
	Total paths	POS <sub>1,2</sub> , % (SE, %)	Total paths	POS <sub>2,3</sub> , % (SE, %)	POS <sub>2,APP</sub> , % (SE, %)	Total paths	POS <sub>3,APP</sub> , % (SE, %)	POS, % (SE, %)
Oncology	17 368	57.6 (0.4)	6533	32.7 (0.6)	6.7 (0.3)	1236	35.5 (1.4)	3.4 (0.2)
Metabolic/ Endocrinology	3589	76.2 (0.7)	2357	59.7 (1.0)	24.1 (0.9)	1101	51.6 (1.5)	19.6 (0.7)
Cardiovascular	2810	73.3 (0.8)	1858	65.7 (1.1)	32.3 (1.1)	964	62.2 (1.6)	25.5 (0.9)
CNS	4924	73.2 (0.6)	3037	51.9 (0.9)	19.5 (0.7)	1156	51.1 (1.5)	15.0 (0.6)
Autoimmune/ Inflammation	5086	69.8 (0.6)	2910	45.7 (0.9)	21.2 (0.8)	969	63.7 (1.5)	15.1 (0.6)
Genitourinary	757	68.7 (1.7)	475	57.1 (2.3)	29.7 (2.1)	212	66.5 (3.2)	21.6 (1.6)
Infectious disease	3963	70.1 (0.7)	2314	58.3 (1.0)	35.1 (1.0)	1078	75.3 (1.3)	25.2 (0.8)
Ophthalmology	674	87.1 (1.3)	461	60.7 (2.3)	33.6 (2.2)	207	74.9 (3.0)	32.6 (2.2)
Vaccines (Infectious Disease)	1869	76.8 (1.0)	1235	58.2 (1.4)	42.1 (1.4)	609	85.4 (1.4)	33.4 (1.2)
Overall	41 040	66.4 (0.2)	21 180	58.3 (2.3)	35.1 (2.2)	7532	59.0 (0.6)	13.8 (0.2)

Lo *et al.* (2018) Estimation of clinical trial success rates and related parameters. *Biostatistics*. p 1–14

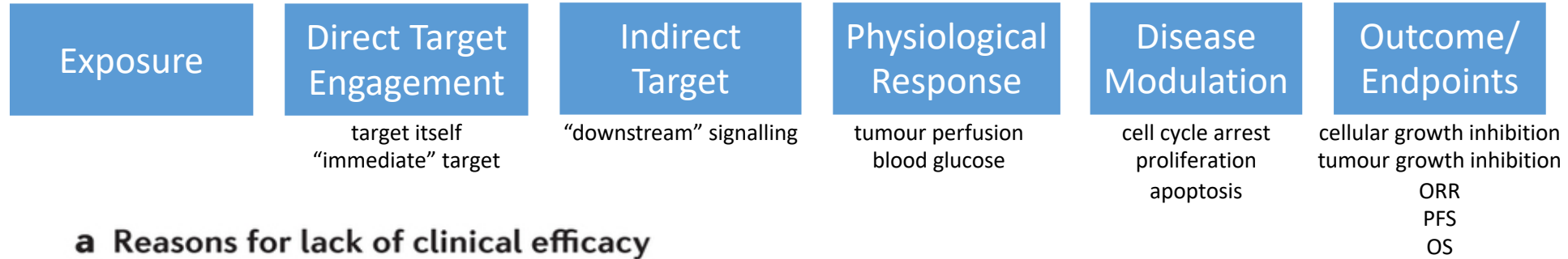
# Pfizer 3 pillars (2012)

- “Pfizer 3 pillars”
  1. Suitable exposure (site of action, duration)
  2. Sufficient target binding
  3. Adequate pharmacology

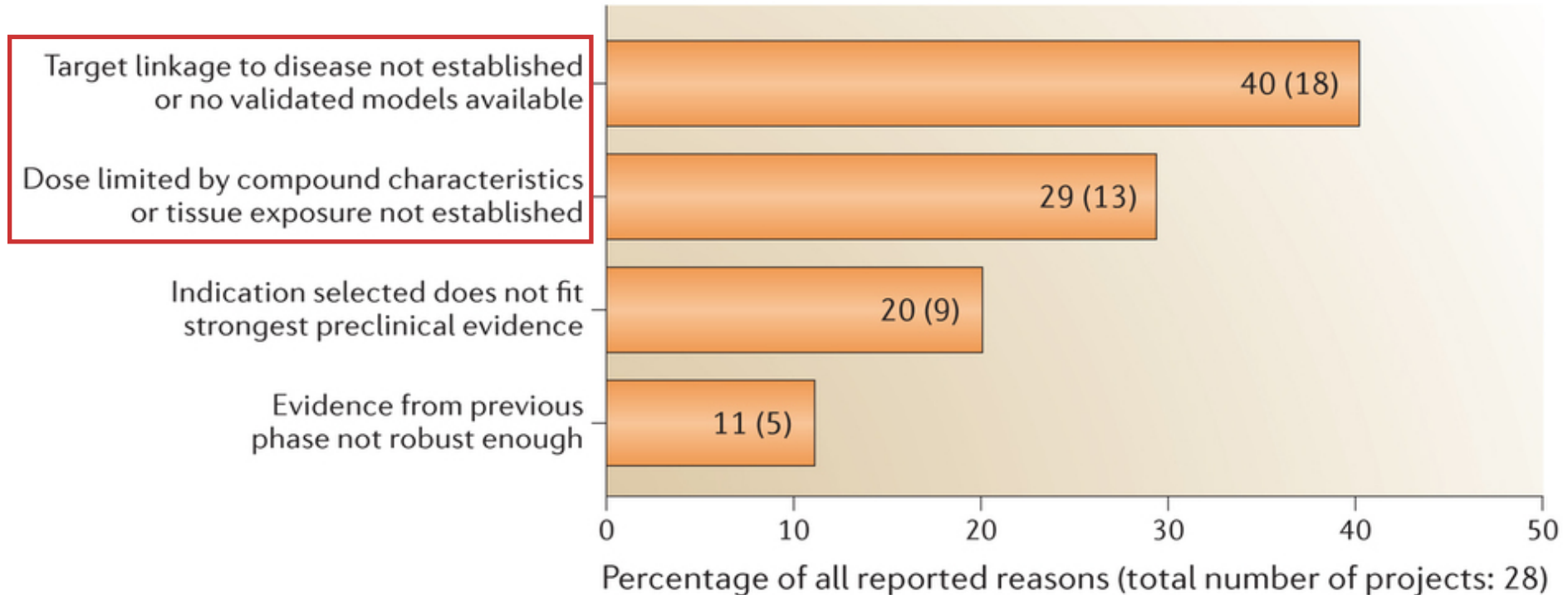


Can the flow of medicines be improved? Fundamental pharmacokinetic and pharmacological principles toward improving Phase II survival *Drug Discovery Today*. 17, 419–424 (2012)

# AstraZeneca: Lack of clinical efficacy (2014)



## a Reasons for lack of clinical efficacy



Lessons learned from the fate of AstraZeneca's drug pipeline: a five-dimensional framework  
*Nature Reviews Drug Discovery* **13**, 419–431 (2014) doi:10.1038/nrd4309

# AstraZeneca 5R's framework

## Right target

- Strong link between target and disease
- Differentiated efficacy
- Available and predictive biomarkers

## Right tissue

- Adequate bioavailability and tissue exposure
- Definition of PD biomarkers
- Clear understanding of preclinical and clinical PK/PD
- Understanding of drug–drug interactions

## Right safety

- Differentiated and clear safety margins
- Understanding of secondary pharmacology risk
- Understanding of reactive metabolites, genotoxicity, drug–drug interactions
- Understanding of target liability

## Right patients

- Identification of the most responsive patient population
- Definition of risk–benefit for given population

## Right commercial potential

- Differentiated value proposition versus future standard of care
- Focus on market access, payer and provider
- Personalized health-care strategy, including diagnostic and biomarkers

“The right culture”

It is vital to ensure that teams are encouraged and rewarded to ask the “killer question”, are recognized for the quality of their science, and are well connected to the external scientific community and supported by experienced leaders with a record of good judgment\*

Lessons learned from the fate of AstraZeneca's drug pipeline: a five-dimensional framework *Nature Reviews Drug Discovery* 13, 419–431 (2014) doi:10.1038/nrd4309

\* Ringel, M., Tollman, P., Hersch, G. & Schulze, U. Does size matter in R&D productivity? If not, what does? *Nature Rev. Drug Discov.* 12, 901–902 (2013).

# The spirit



“We can judge our progress by the courage of our questions and the depth of our answers, our willingness to embrace what is true rather than what feels good.”

— Carl Sagan (1934-1996)

# THE OPEN PROJECT (TOP)

- Vision

Affordable and effective novel therapies discovered and developed based on all accessible, relevant data in a timely manner

- Mission

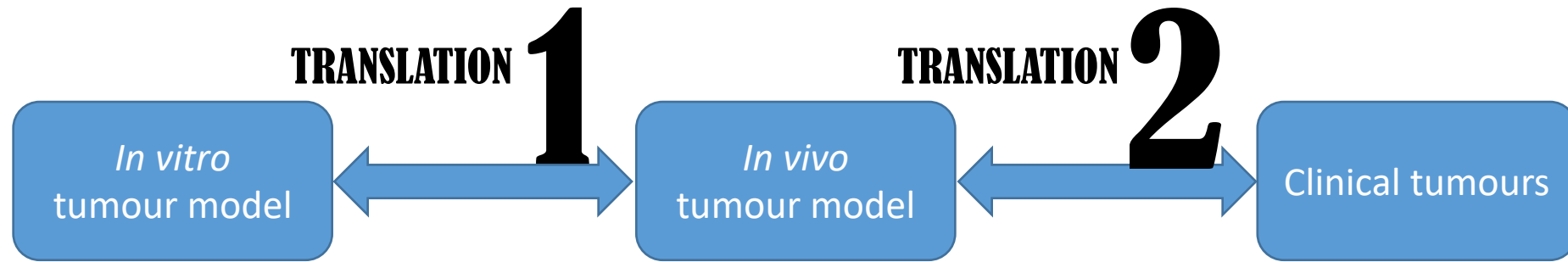
Pioneer in translational modelling to develop, validate and improve quantitative methods and tools for accurate experimental design to enable robust decision making in drug discovery and development

# THE OPEN PROJECT (TOP)

- Participants
  - Open: Any one can join for free to share data, models, codes and ideas
  - Transparent: All results are properly documented to help the community
  - Meritocracy: Participants are required to demonstrate understanding of the code, rules, and culture of the project before being invited to join



# Translational Oncology Modelling Goals



## Challenges

Translation 1. *in vitro* efficacy <-> *in vivo* efficacy

- How do *in vitro* tumour models grow?
- How do *in vivo* tumour models grow?
  - Which rate law accurately recapitulates tumour growth?
  - Which rate law accurately predicts tumour growth?
- Can *in vivo* gene expression predict *in vivo* tumour growth?

Growth: Exchangeable with “response to treatments”

# 1. *In vitro* data -> *In vivo* design

	Exposure	Target Engagement	Disease Modulation	Outcome
<i>In vitro</i>	Medium	Direct / Indirect	DM	Efficacy
<i>In vivo</i>	Plasma Tissue	Direct / Indirect	DM	Efficacy
Clinical	Plasma	Direct / Indirect	DM	Efficacy

Green double-headed arrows connect adjacent cells in the *In vitro* and *In vivo* rows. A green arrow with a '1' in a circle points from the *In vitro* Efficacy cell to the *In vivo* Efficacy cell.

**Objective:** Define the necessary compound profile to fulfill PoM criterion

## Value

- Project direction: A valid target compound profile to support PoM study
- Time saving: Shorter path to an *in vivo* experiment that demonstrates understanding of efficacy

## Deliverables

- Target compound profile criteria
- Design of an *in vivo* experiment to demonstrate *in vivo* efficacy

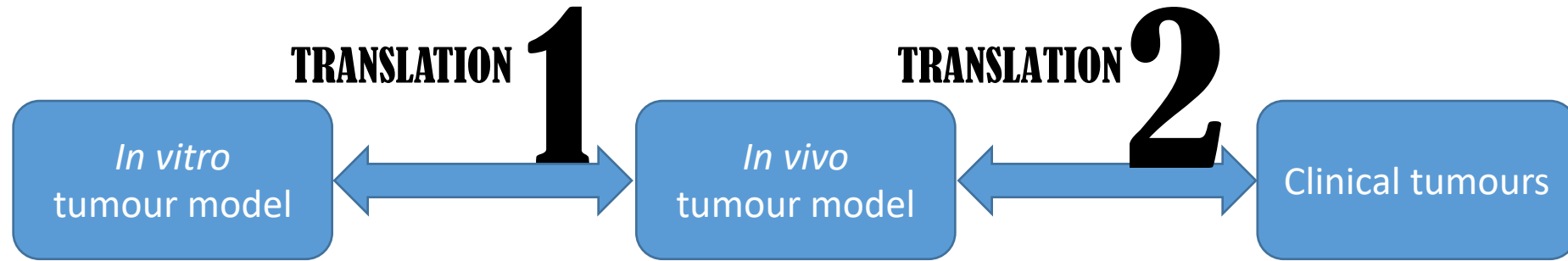
## Information required

- *In vitro* Target Engagement markers
- *In vitro* Disease Modulation marker
- Modelling that predicts *in vivo* efficacy based on *in vitro* efficacy

## Triggering questions

- How would you forecast *in vivo* efficacy based on IC<sub>50</sub> (*in vitro* potency)?
- Do your data confirm the therapeutic concept?

# Translational Oncology Modelling Goals



## Challenges

Translation 2. *in vivo* efficacy <-> clinical efficacy

- How do clinical tumours respond?
- Can *in vivo* tumour models forecast clinical tumour responses?

# 2. Clinical data -> *In vivo* design

	Exposure	Target Engagement	Disease Modulation	Outcome
<i>In vitro</i>	Medium	Direct / Indirect	DM	Efficacy
<i>In vivo</i>	Plasma Tissue	Direct / Indirect	DM	Efficacy
Clinical	Plasma	Direct / Indirect	DM	Efficacy

Diagram annotations: A green arrow points from 'Plasma Tissue' to 'Plasma'. Green double-headed arrows connect 'Plasma Tissue' to 'Direct / Indirect', 'Direct / Indirect' to 'DM', and 'DM' to 'Efficacy'. A green circle with the number '2' is placed over the 'Efficacy' cell in the 'Clinical' row, with a green arrow pointing up from it to the 'Efficacy' cell in the 'In vivo' row and a green arrow pointing down from it to the 'Efficacy' cell in the 'Clinical' row.

**Objective:** Define and support PoC strategy (phases 1-2)

### Value

- Project direction: A valid target compound profile supported by clinical evidence
- Time saving: Shorter path to an *in vivo* experiment supporting clinical feasibility

### Deliverables

- Identify clinical efficacious doses and optimal dosing schedule for the (combination) treatment
- Identify the best clinical combination partner

### Information required

- Competitor/combination compounds
  - PK/PD information of tumour models
  - Clinical popPK
- Own compound
  - Preclinical PK/PD/Efficacy data
  - Forecasted clinical PK

# 3. *In vivo* data -> Clinical design

	Exposure	Target Engagement	Disease Modulation	Outcome
<i>In vitro</i>	Medium	Direct / Indirect	DM	Efficacy
<i>In vivo</i>	Plasma Tissue	Direct / Indirect	DM	Efficacy
Clinical	Plasma	Direct / Indirect	DM	Efficacy

Diagram annotations: Green double-headed arrows connect adjacent cells in each row. A green arrow points from 'Efficacy' in the *In vitro* row to 'Efficacy' in the *In vivo* row, with a circled '1' next to it. A green arrow points from 'Plasma' in the *In vivo* row to 'Plasma' in the Clinical row, with a circled '3' next to it. A green arrow points from 'Direct / Indirect' in the *In vivo* row to 'Direct / Indirect' in the Clinical row.

**Objectives:** Define PoP criterion (phase 1)

## Benefit

- Project direction: A valid clinical biomarker experiment design supported by preclinical science
- Time saving: The right experiment indicating signs of clinical efficacy at the first time

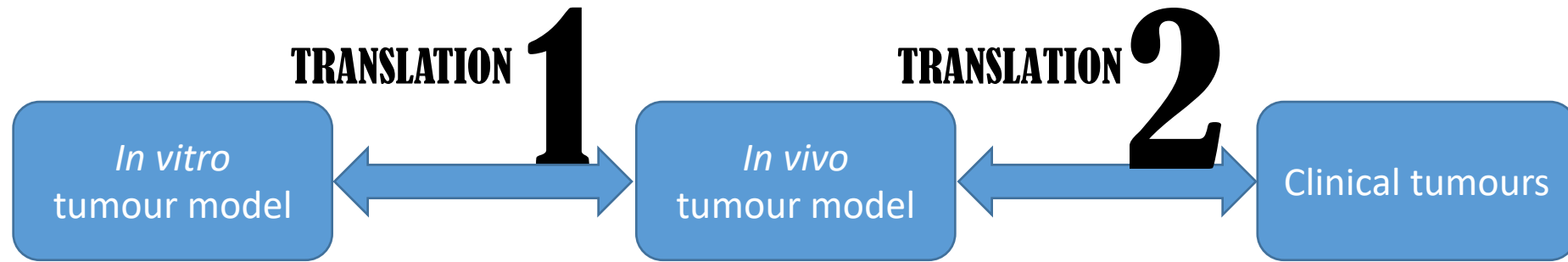
## Deliverables

- Forecast clinical PK
- Target clinical modulation of TE (i.e. minimum TE that is needed to modulate disease)
- Recommended timing of biopsy to assess TE

## Information required

- *In vitro* and *in vivo* Target Engagement marker
- *In vitro* and *in vivo* Disease Modulation marker
- Preclinical TGI data
- Clinical tumour imaging data (RECIST criteria)
- Forecast clinical PK

# Translational Oncology Modelling Goals



## Challenges we tackle here

- How do *in vivo* tumour models grow?
  - Which rate law accurately recapitulates tumour growth?
  - Which rate law accurately predicts tumour growth?
- Can *in vivo* gene expression predict *in vivo* tumour growth?

## 2. *IN VIVO* TUMOUR GROWTH RATE LAWS

- Which rate law accurately recapitulates tumour growth?
- Which rate law accurately predicts tumour growth?

# THE BACKGROUND



# Tumour Growth Rate Laws

- Linear
  - Tumour radius expands linearly
  - TV is used for fitting for consistent error model:  $TV = \frac{4}{3}\pi(r_0 + g * t)^3$
- Exponential
  - $TV = TV_0 e^{at}$
- Exponential-linear: incompatible with tumour that shrinks
  - $\frac{dV}{dt} = a_0 V, t \leq \tau; \frac{dV}{dt} = a_1, t > \tau$ . V is smooth:  $\tau = \frac{1}{a_0} \log \left( \frac{a_1}{a_0 V_0} \right)$
- Logistic
  - $\frac{dV}{dt} = aV \left( 1 - \frac{V}{K} \right)$   $a$ : growth rate (/day);  $K$ : carrying capacity (mm<sup>3</sup>)
- Gompertz
  - $\frac{dV}{dt} = \alpha e^{-\beta t} V$   $\alpha$ : growth rate (/day);  $\beta$ : decay rate (/day)

# Linear (actually cubic) growth

- Tumour radius expands linearly:  $TV = \frac{4}{3}\pi(r_0 + g * t)^3$
- First proposed by Mayneord in 1932
- Assumption: Solid tumour growth is limited to a thin surface layer of cells (competition for space, not nutrients)
- Notable data that “support” the model:
  - Jensen’s rat sarcoma: n=8
  - Walker rat tumour: n=5
  - *In vitro* cell colonies with proliferating rim: n=15
  - Cell line derived xenografts: n=16
  - Clinical untreated low-grade gliomas: n=27

Mayneord, W.V. (1932) On a law of growth of Jensen’s rat sarcoma. *Am. J. Cancer* 16, 841–846

Schrek, R. (1935) A quantitative study of the growth of the Walker rat tumor and the Flexner–Jobling rat carcinoma. *Am. J. Cancer* 24, 807–822

Brú, A. *et al.* (2003) The universal dynamics of tumor growth. *Biophys. J.* 85, 2948–2961

Mandonnet, E. *et al.* (2003) Continuous growth of mean tumor diameter in a subset of grade II gliomas. *Ann. Neurol.* 53, 524–528

# Exponential growth

- $TV = TV_0 e^{at}$  with doubling time  $\ln(2)/a$
- First proposed by Collins in 1956
- Assumption: Tumour cell doubling time is constant
- Notable clinical data that “support” the model:
  - Soft tumour: L1210 leukemia in 1970
  - Solid tumours (sometime restricted to the initial period):
    - Untreated breast cancer: n=12
    - Primary pulmonary malignancies: n=41
    - Pulmonary metastases from various primary tumours: n=118
    - Pulmonary metastases from mesenchymal malignancies: n=16
    - Renal cell carcinomas: n=16

Collins, V.P. *et al.* (1956) Observations on growth rates of human tumors. *Am. J. Roentgenol. Radium Ther. Nucl. Med.* 76, 988–1000

Shackney, S.E. (1970) A computer model for tumor growth and chemotherapy, and its application to L1210 leukemia treated with cytosine arabinoside (nsc-63878). *Cancer Chemother. Rep.* 54, 399–429

Friberg, S. and Mattson, S. (1997) On the growth rates of human malignant tumors: implications for medical decision making. *J. Surg. Oncol.* 65, 284–297

# Exponential-linear

- Exponential-linear: incompatible with tumour that shrinks
  - $\frac{dV}{dt} = a_0V, t \leq \tau; \frac{dV}{dt} = a_1, t > \tau$ .  $V$  is smooth:  $\tau = \frac{1}{a_0} \log \left( \frac{a_1}{a_0V_0} \right)$
  - Simeoni model approximates the original model

$$\frac{dw(t)}{dt} = \frac{\lambda_0 \cdot w(t)}{\left[ 1 + \left( \frac{\lambda_0}{\lambda_1} \cdot w(t) \right)^\Psi \right]^{1/\Psi}}$$

$$w(0) = w_0$$

$$\Psi = 20$$

- Popular among PK/PD community
- Notable data that “support” the model:
  - Various types of CDX tumour: n=30

Simeoni, M *et al.* (2004) Pharmacokinetic-Pharmacodynamic Modeling of Tumor Growth Kinetics in Xenograft Models after Administration of Anticancer Agents. *Cancer Research*. 64, 1094 – 1101

# Logistic growth

- $\frac{dV}{dt} = aV \left(1 - \frac{V}{K}\right)$   $a$ : growth rate (/day);  $K$ : carrying capacity ( $\text{mm}^3$ )
- First proposed by Robertson in 1923
- Assumption: Solid tumour growth is limited to a finite size
- Notable data that “support” the model:
  - Breast cancer:  $n=433$

Robertson, T.B. (1923) The Chemical Basis of Growth and Senescence. J.B. Lippincott & Co.

# Gompertz growth

- $\frac{dV}{dt} = \alpha e^{-\beta t} V$   $\alpha$ : growth rate (/day);  $\beta$ : decay rate (/day)
- First proposed by Casey in 1934
- Assumption: Diffusion and competition for nutrients / tumour self-seeding
- Notable data that “support” the model:
  - *In vivo* animal tumours: n=19
  - B-16 melanoma: n=8; 13762 carcinoma: n=10
  - Breast cancer patients: n=250

Klein, G. and Revesz, L. (1953) Quantitative studies on the multiplication of neoplastic cells in vivo. I. growth curves of the Ehrlich and mc1m ascites tumors. *J. Natl. Cancer Inst.* 14, 229–277

Norton L *et al.* (1976) Predicting the course of Gompertzian growth. *Nature* **264**: 542–545

Norton, L. (1988) A Gompertzian model of human breast cancer growth. *Cancer Res.* 48, 7067–7071

# Exceptions

- Spontaneous regression
- No growth
- Irregular growth rates
- Accelerations of growth

Friberg, S. and Mattson, S. (1997) On the growth rates of human malignant tumors: implications for medical decision making. *J. Surg. Oncol.* 65, 284–297

# Goals

- Which rate laws accurately recapitulate tumour growth?
  - Parametric inference: DATA + MODEL STRUCTURE → PARAMETRIC DISTRIBUTION
  - Interpolation: Models are often used to make interpolated predictions
- Which rate laws accurately predict tumour growth?
  - Extrapolation: future tumour growth
  - Good inference is necessary but not sufficient for good prediction
  - Uncertainty in model structure



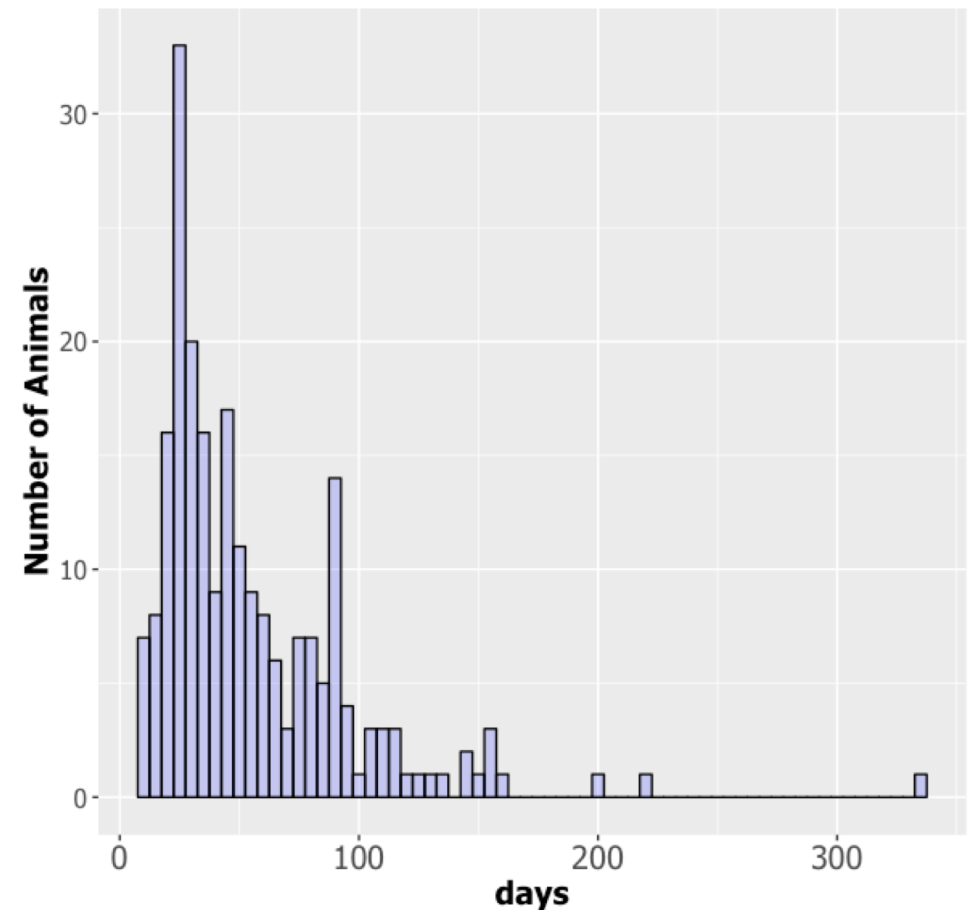
# THE DATA

# Novartis Mouse Clinical Trial

## PDX control experiment overview

- 224 PDX's in total
- Most control experiments are less than 100 days
- First peak: 15-20 days
- Second peak: 85-90 days

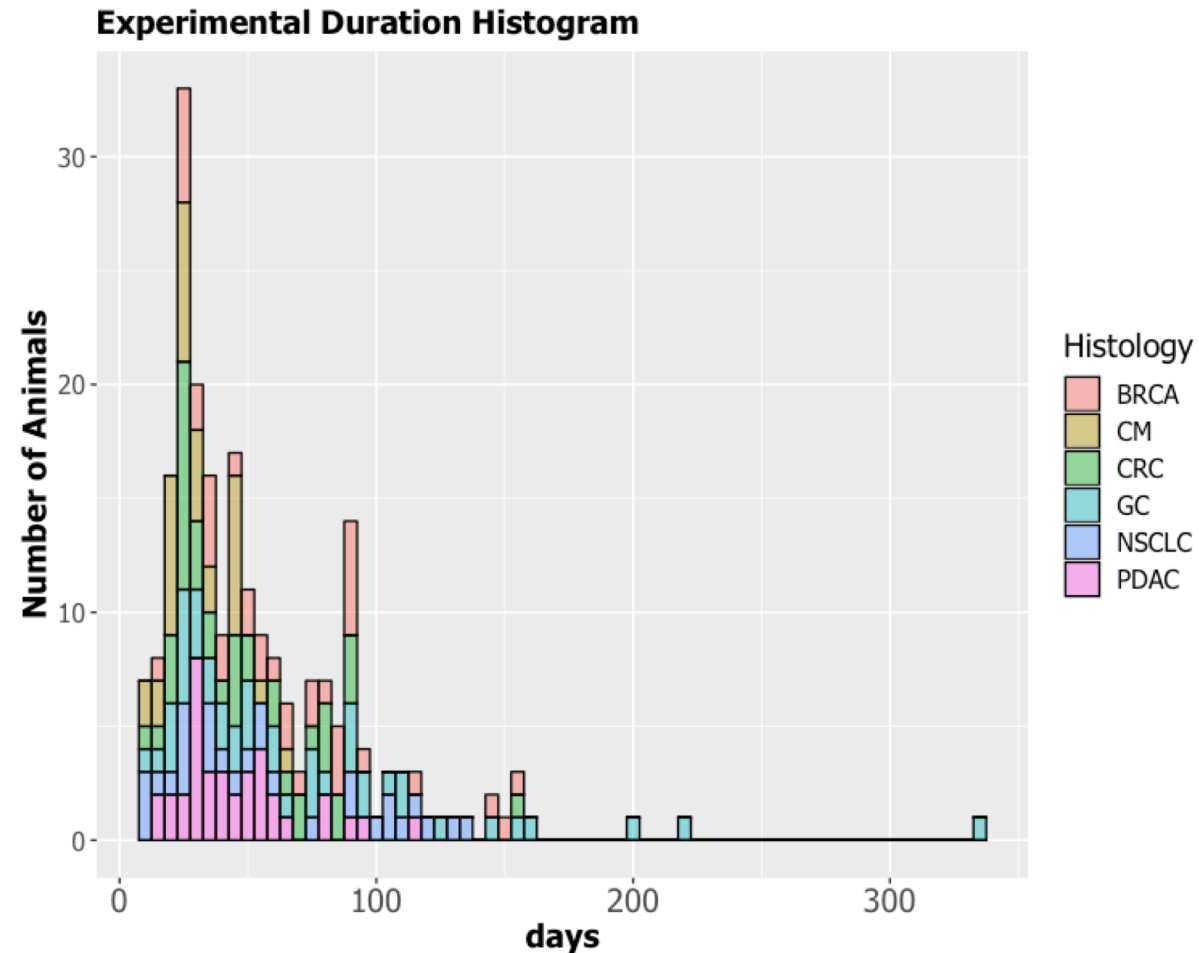
Experimental Duration Histogram



# Novartis Mouse Clinical Trial

## PDX control experiment by histology

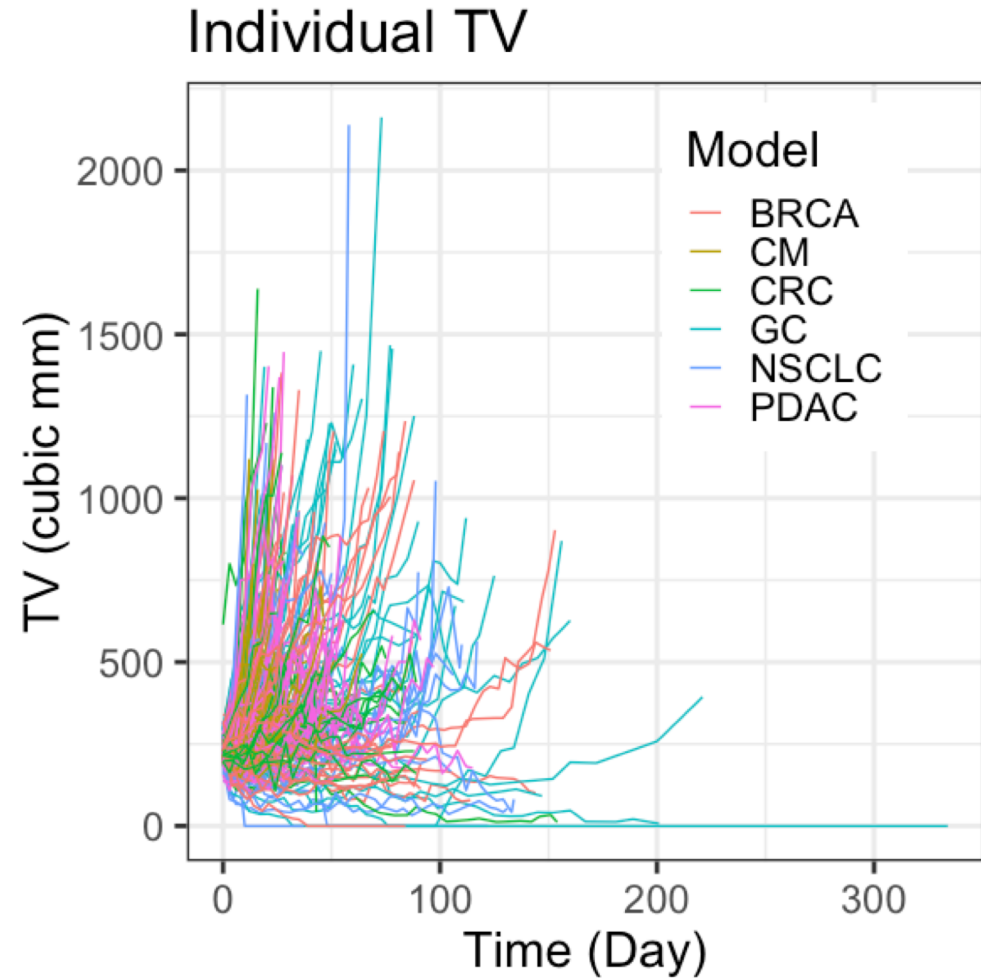
- Histology
  - GC (gastric) 44
  - CRC (colorectal) 42
  - BRCA (breast) 39
  - NSCLC (non-small cell lung) 29
  - PDAC (panc duct adeno) 37
  - CM (cutaneous melanoma) 33
- Experimental duration distribution
  - Similar across histology
- Second peak (85-90 days):
  - BRCA
  - CRC
  - GC
- All CM experiments  $\leq 64$  days



# Novartis Mouse Clinical Trial

## PDX control experiment overview

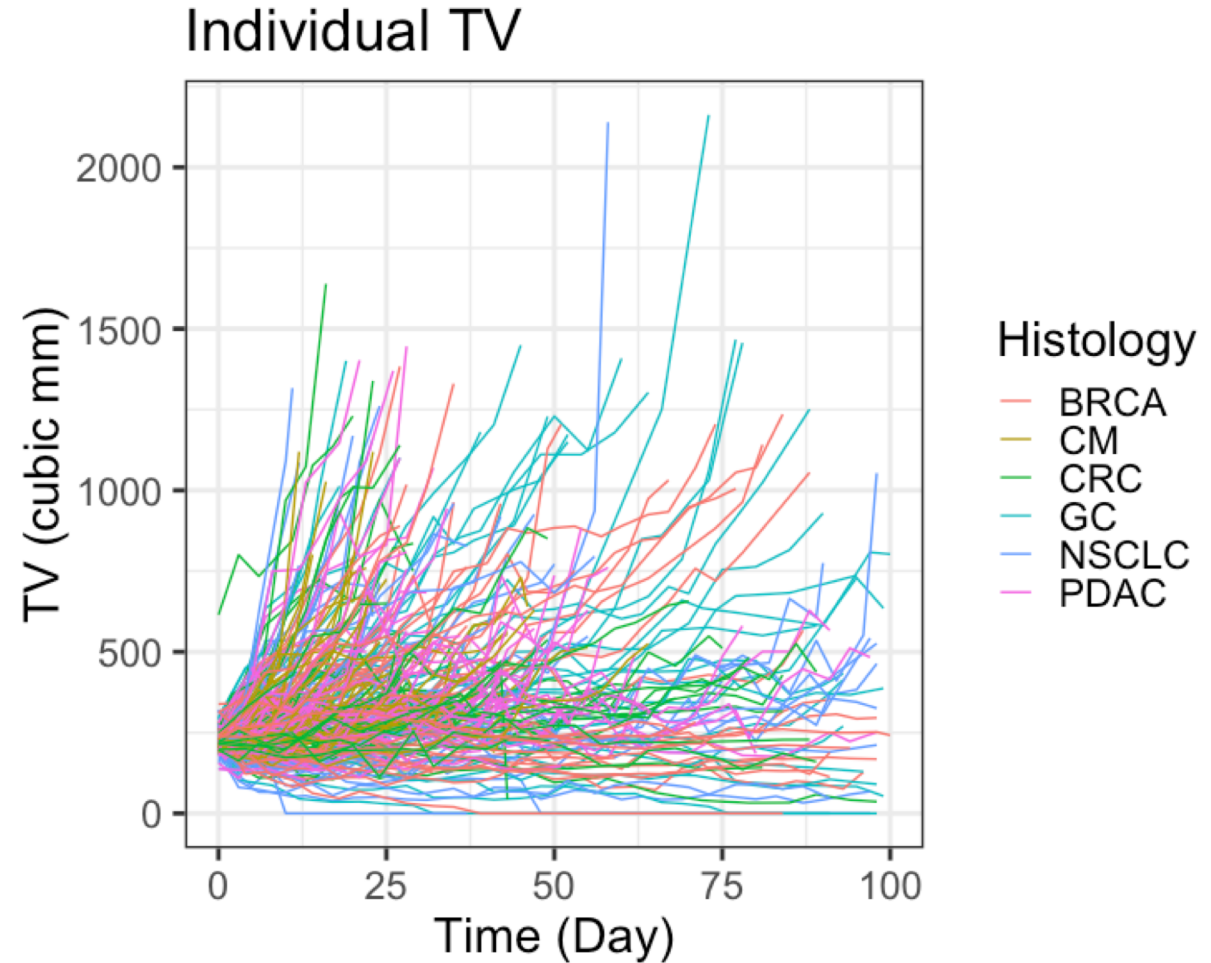
- Starting volume
  - 100~300mm<sup>3</sup>
- Exceptions
  - Spontaneous regression: Yes
  - No growth: Yes
  - Irregular growth rates: Maybe
  - Accelerations of growth: Morphological or genetic adaptation?



# Novartis Mouse Clinical Trial

## PDX control experiment overview

- Growth (day 0~100)
  - Tumour growth varies by histology?
  - Hard to tell from this plot
  - See each histology next





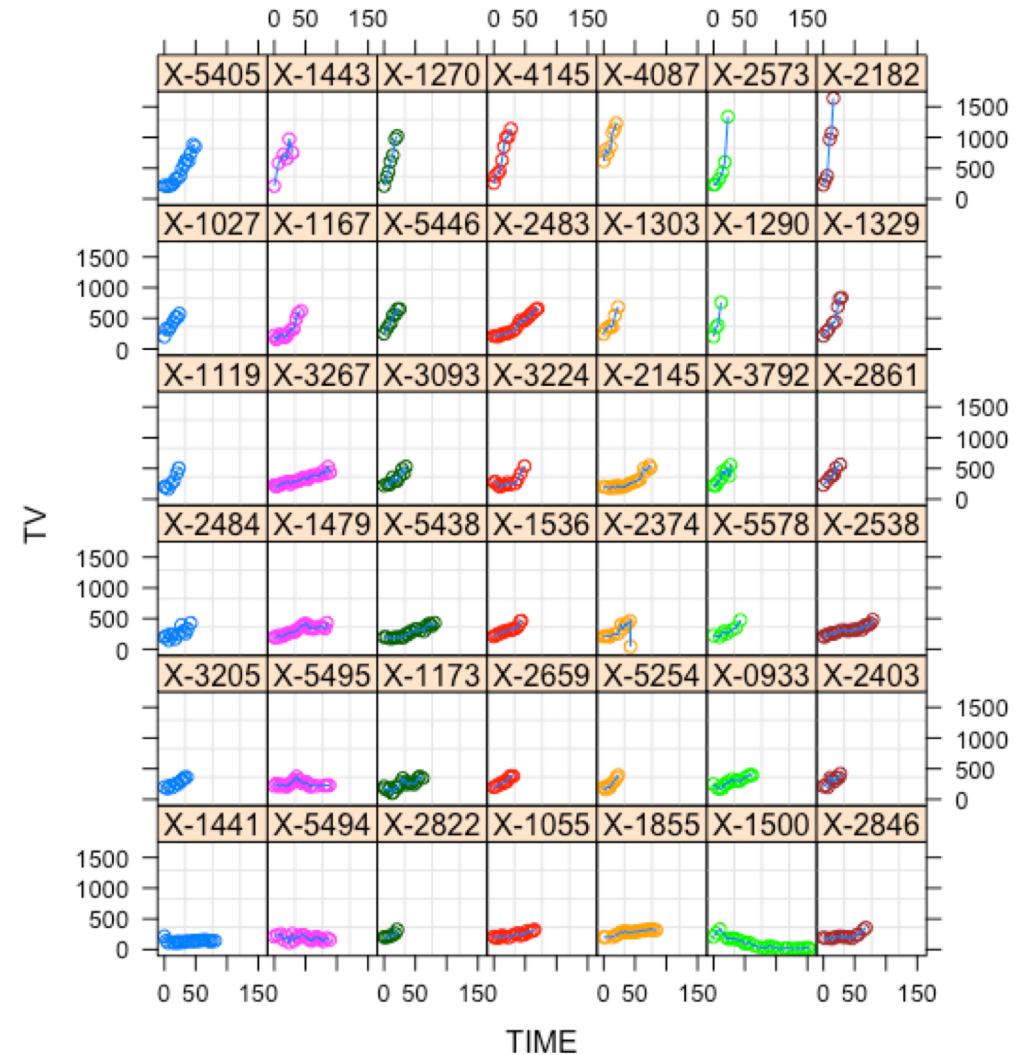
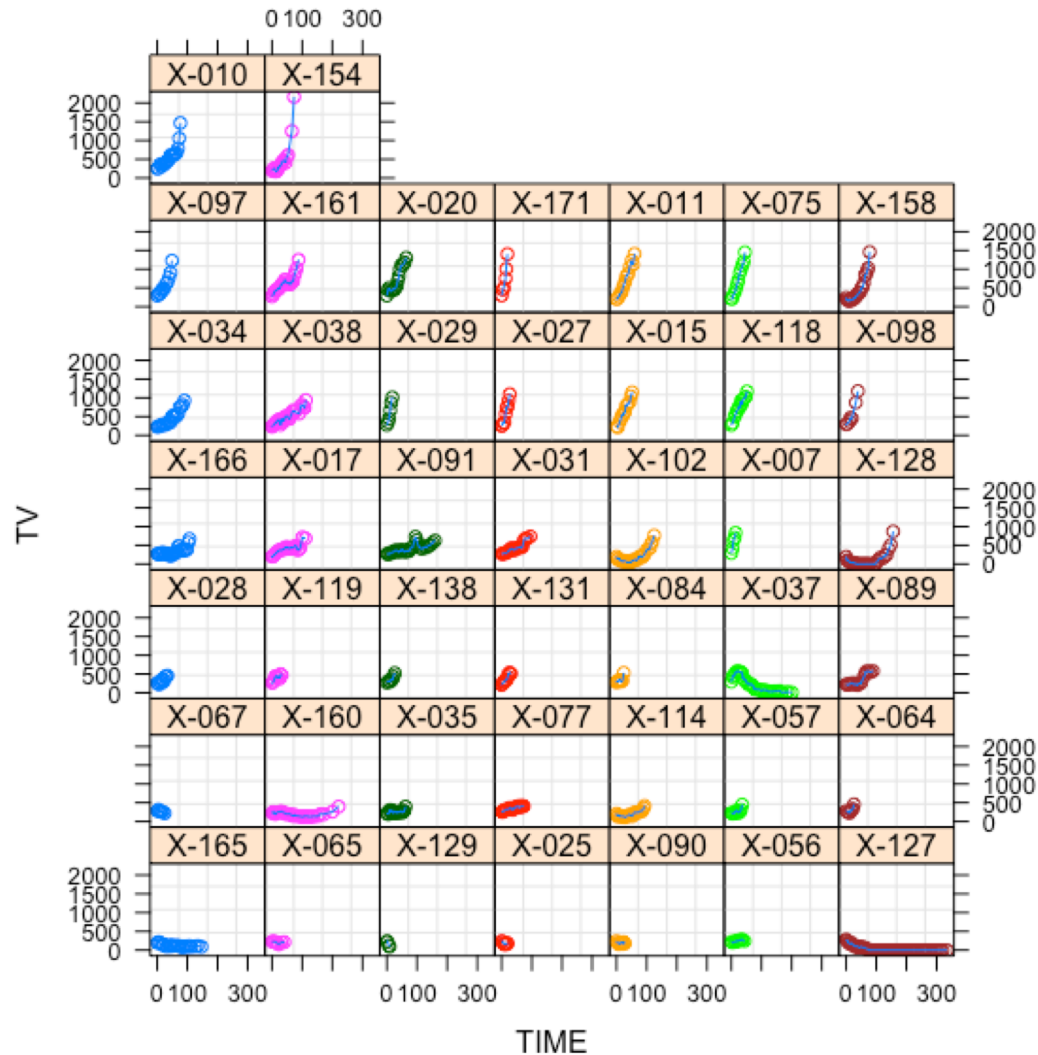
# PDX control experiments



X: PDX number

GC

CRC

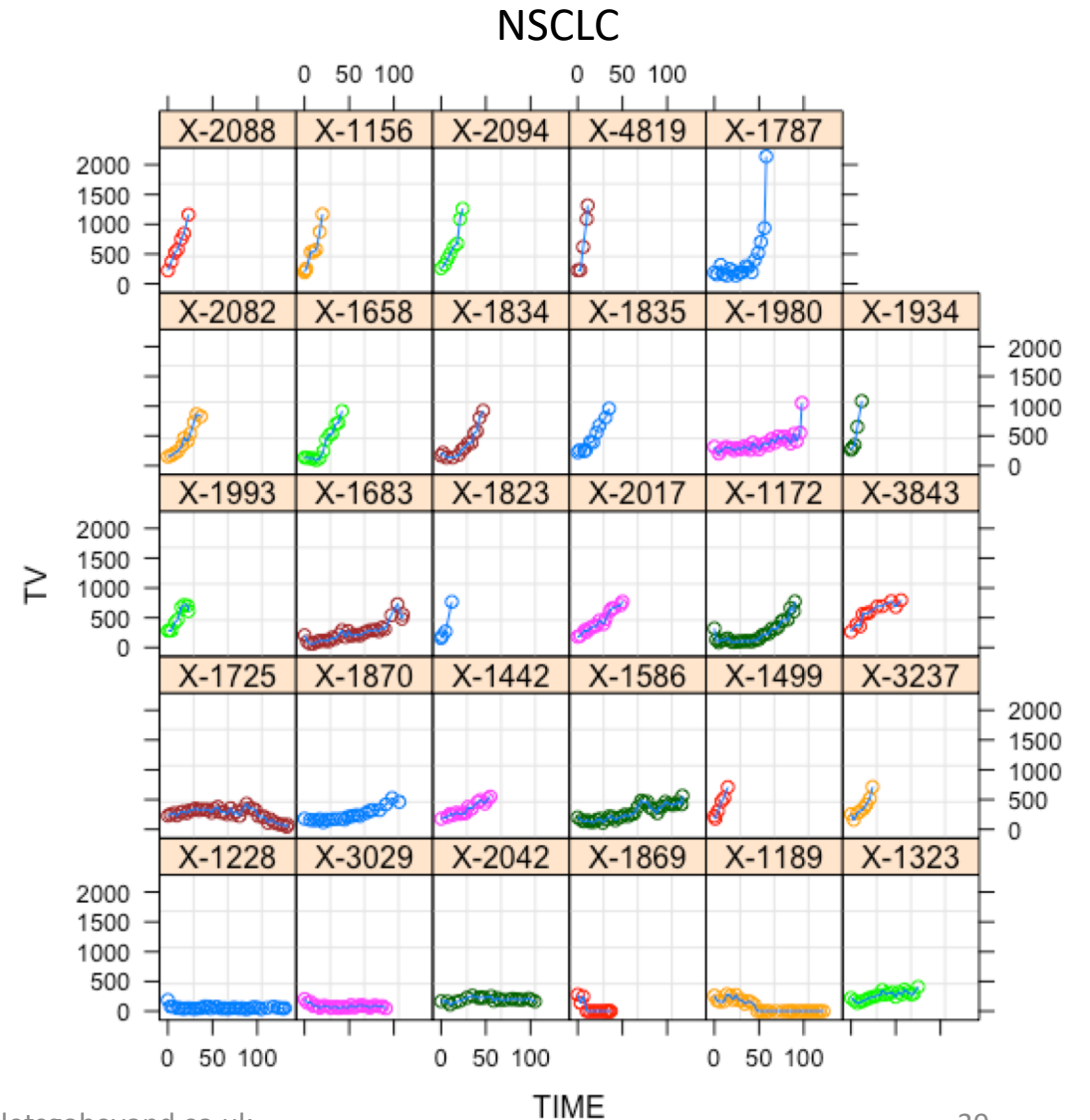
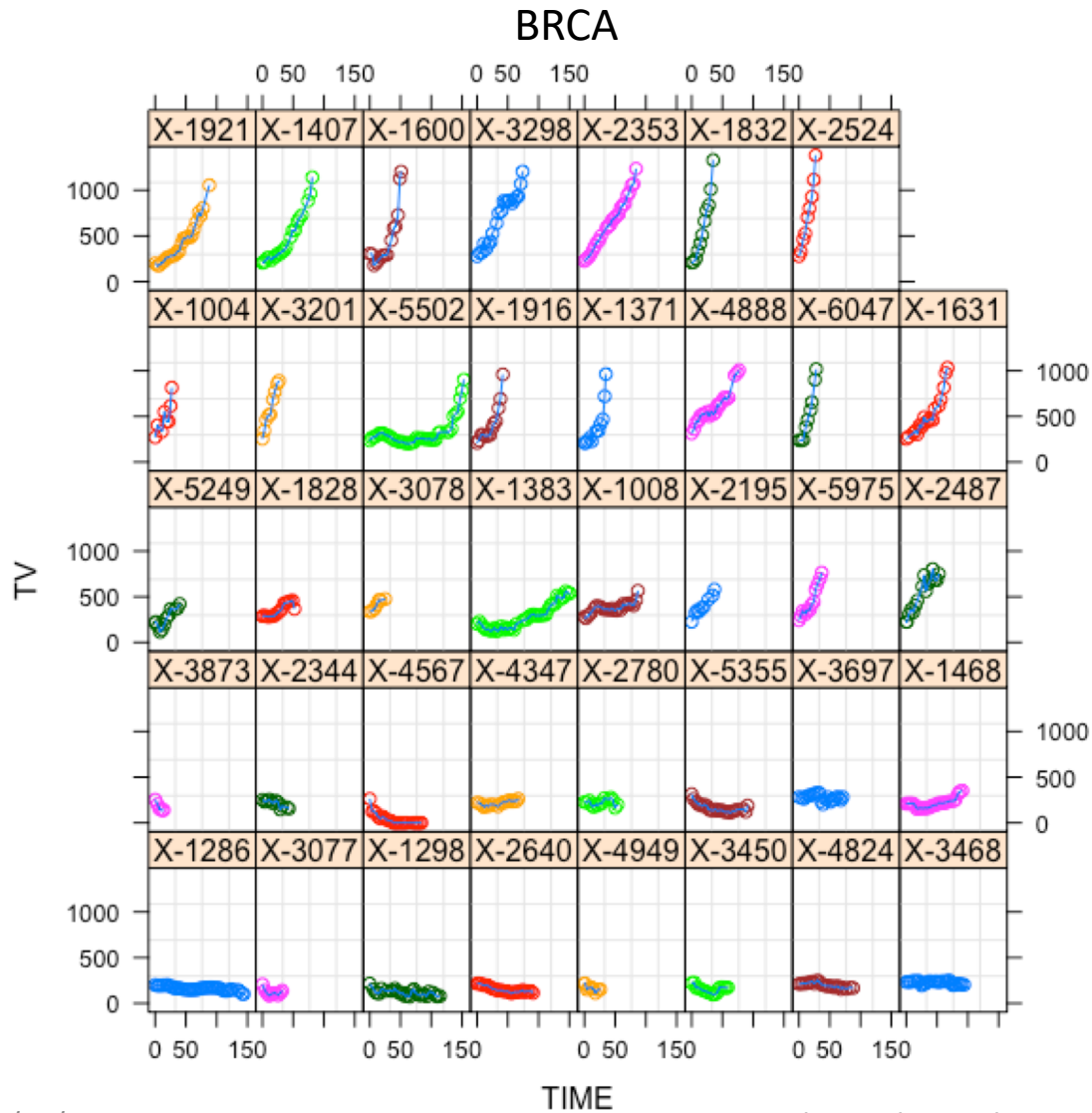




# PDX control experiments



X: PDX number



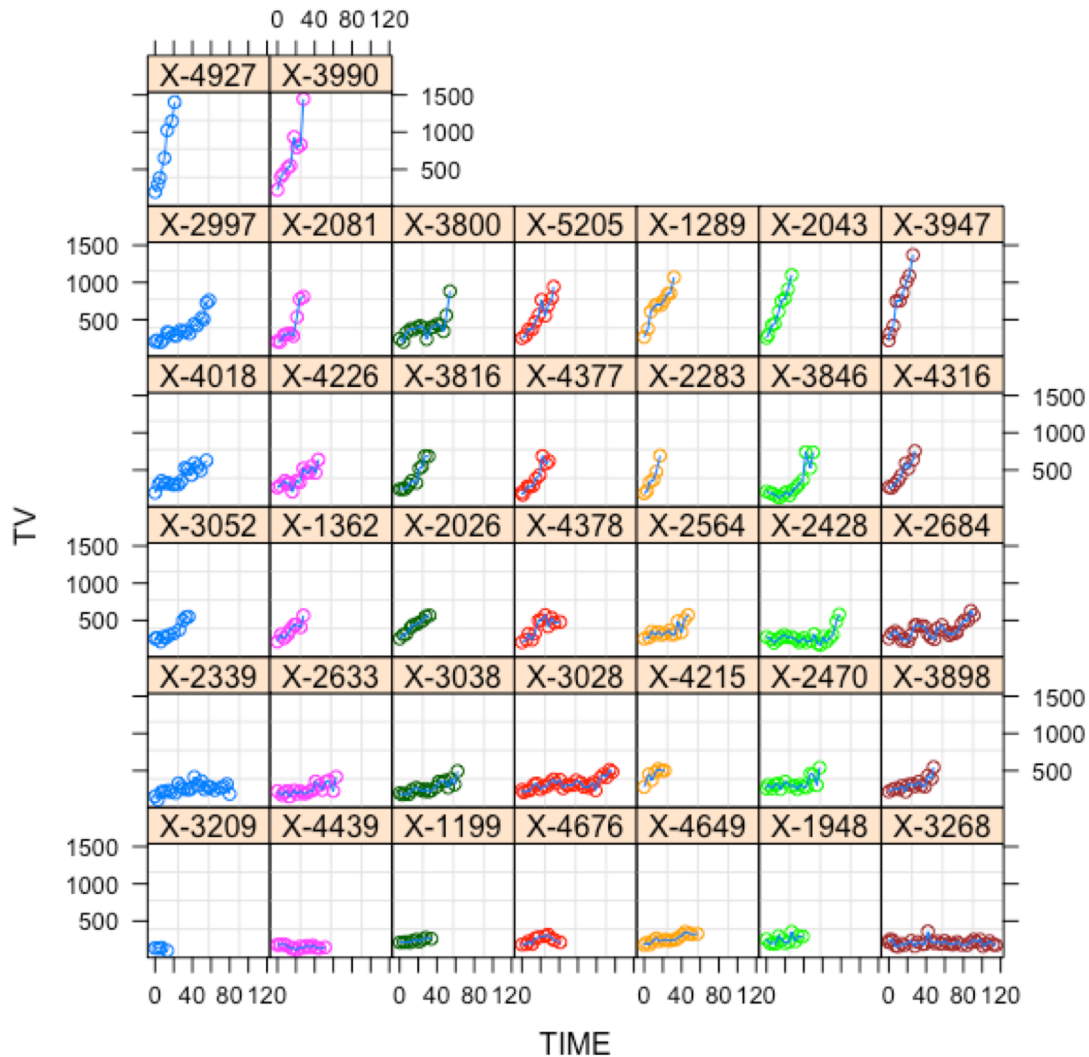


# PDX control experiments

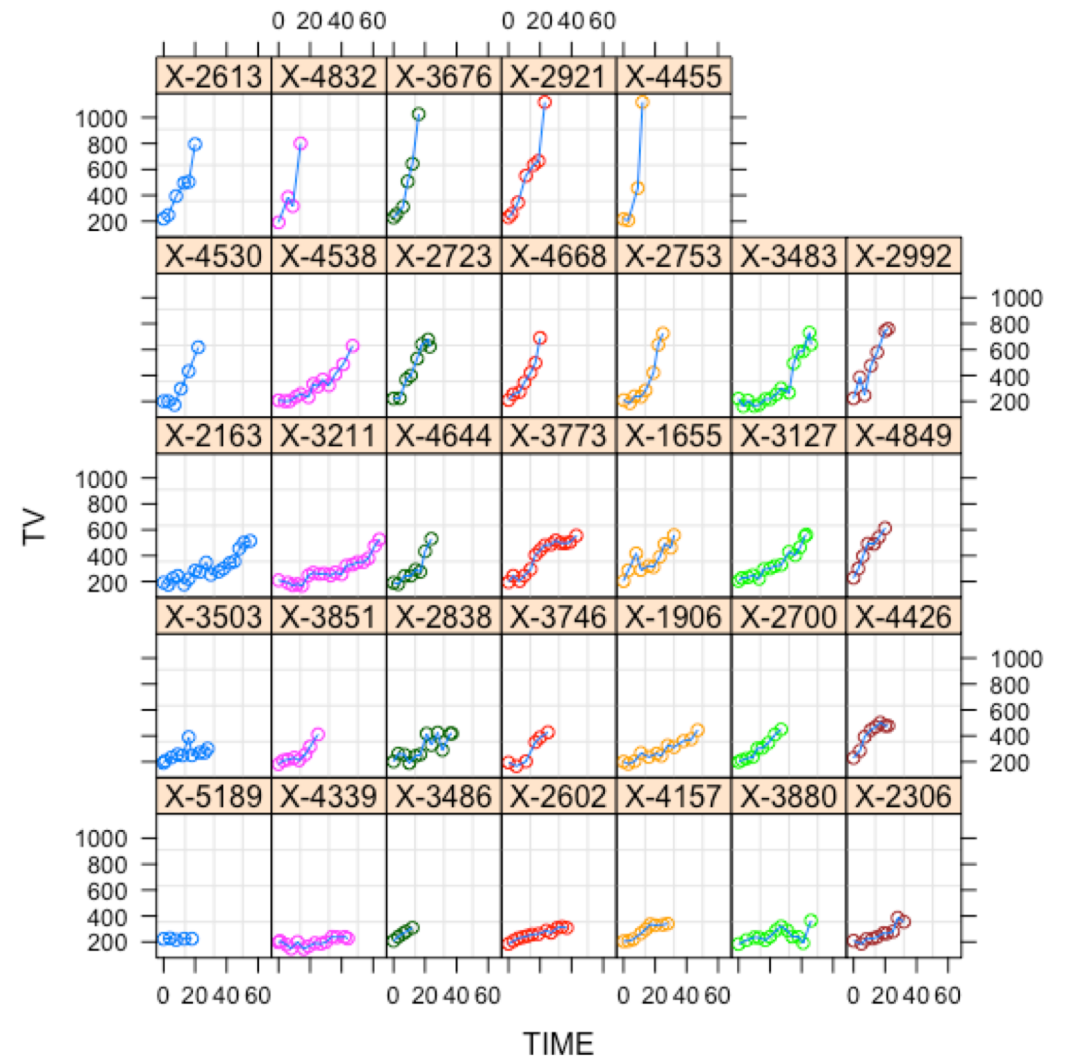


X: PDX number

PDAC



CM





# WHICH RATE LAW ACCURATELY RECAPITULATES TUMOUR GROWTH?

# Linear model fits well

- Fixed:  $r_0$ ,  $g$ ; Random:  $\text{diag}(r_0, g)$

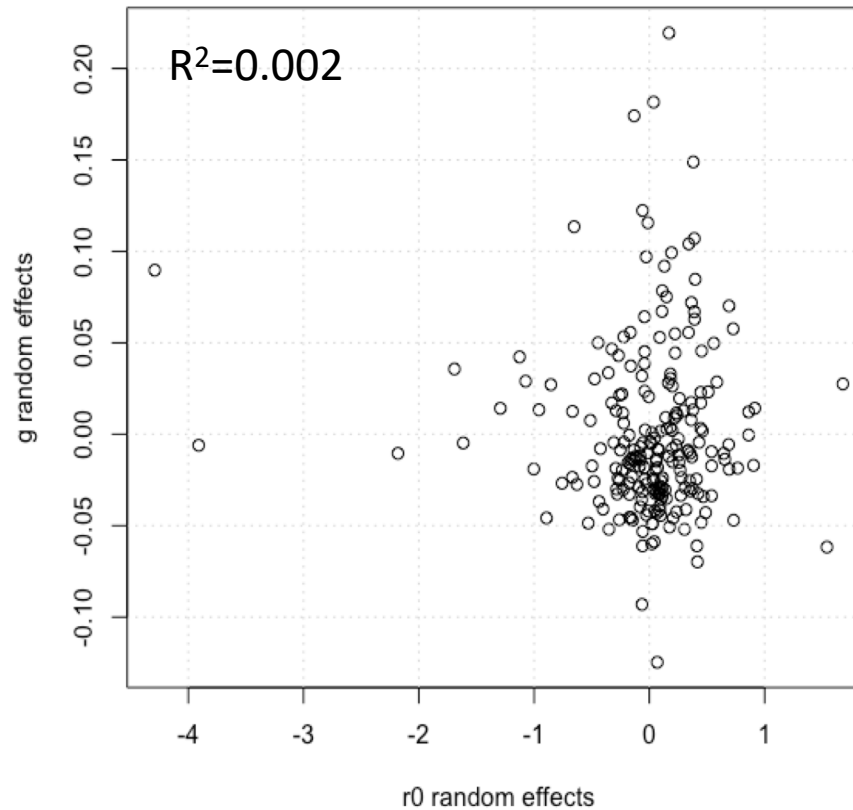
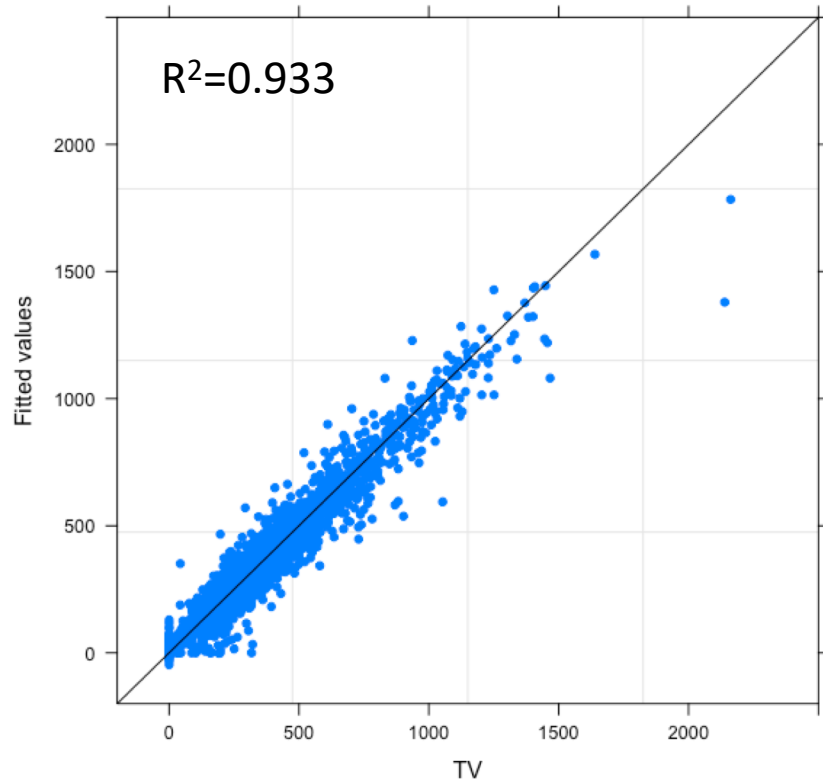
	Value	Std. Error	DoF	t-value	P-value
$r_0$	3.6	0.043	3160	84	0
$g$	0.041	0.0032	3160	13	0

Linear Model

$$TV = \frac{4}{3} \pi (r_0 + g * t)^3$$

Fitting by NLME in R:  
 $TV_0 = 200 \text{ mm}^3$   
 $g = 0.041 \text{ mm/day}$

Linear Model



# Exponential model fits well

- Fixed:  $TV_0, a$ ; Random:  $\text{diag}(TV_0, a)$

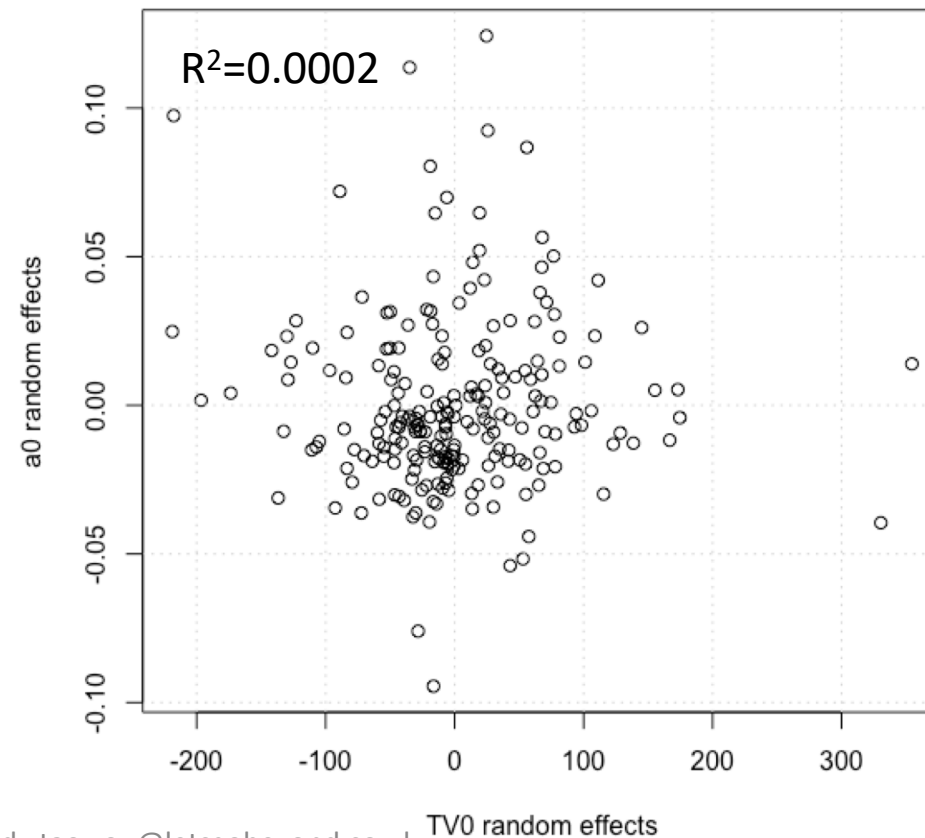
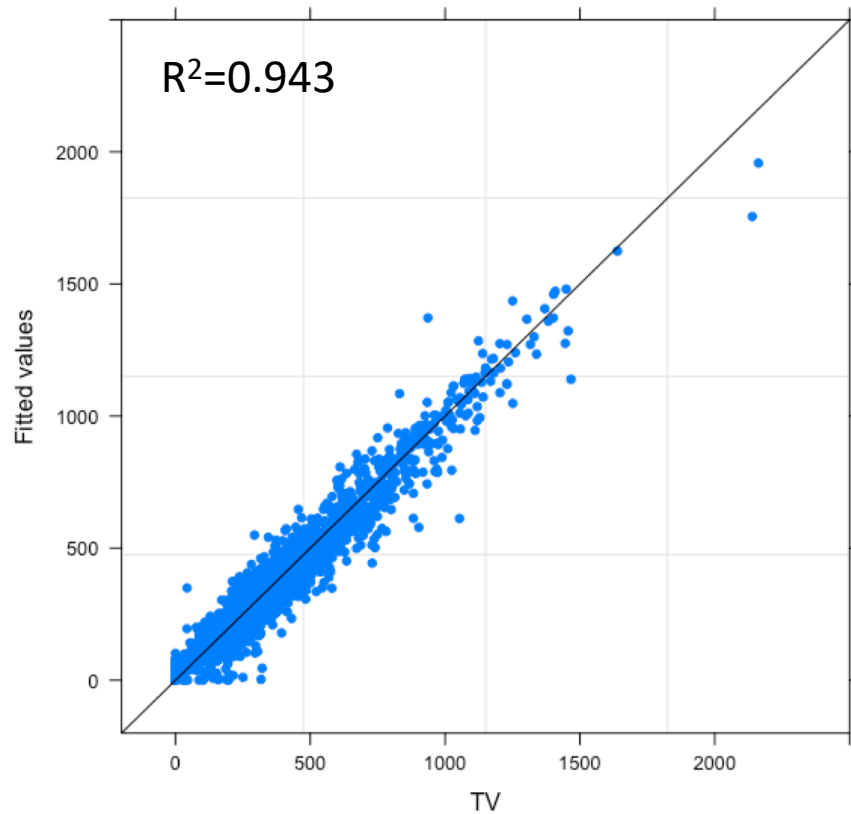
$$TV = TV_0 e^{at}$$

	Value	Std. Error	DoF	t-value	P-value
$TV_0$	220	5.4	3160	41	0
$a$	0.026	0.0020	3160	13	0

Fitting by NLME in R:  
 $TV_0 = 220 \text{ mm}^3$   
 $a = 0.026$

Exponential Model

Exponential Model



# Exponential-linear model was actually just exponential

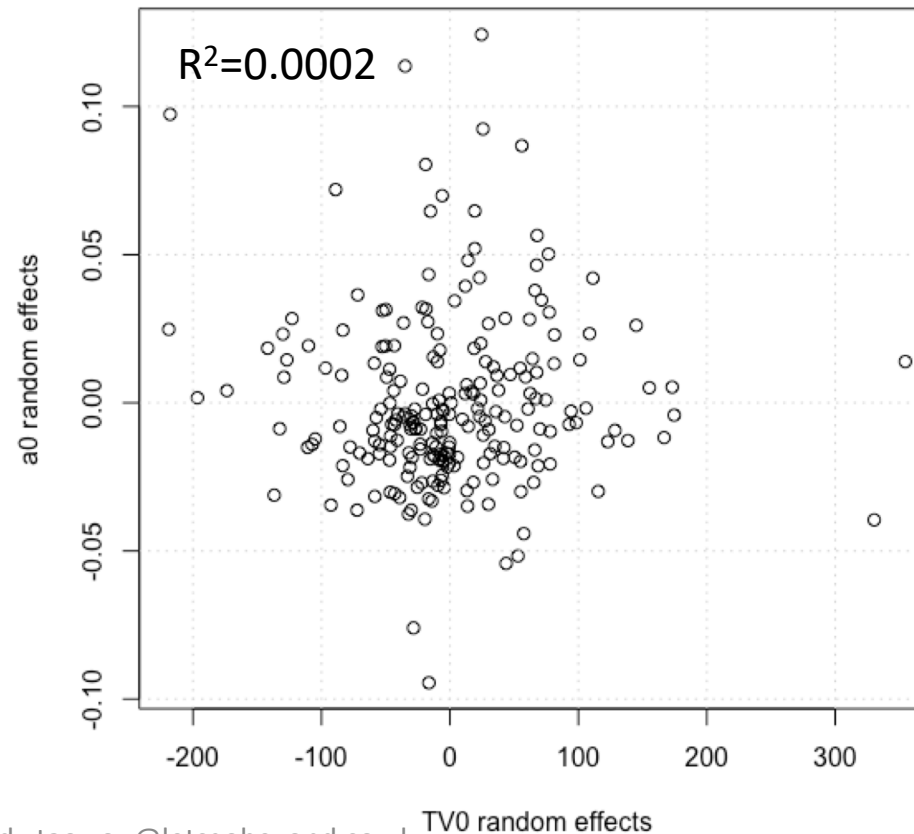
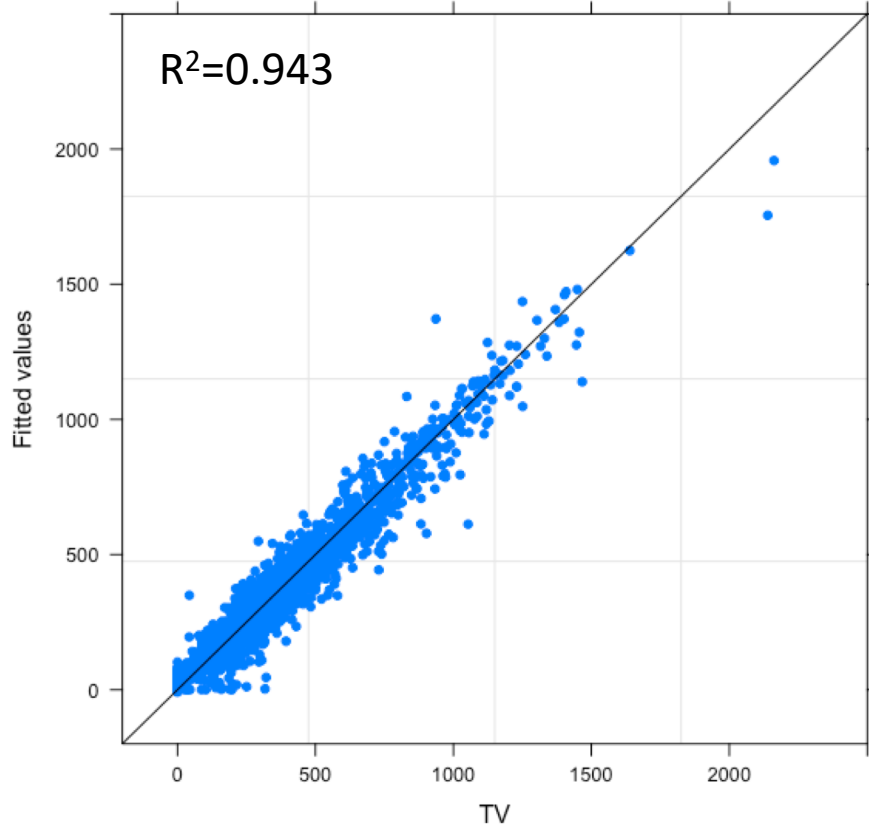
- Fixed:  $TV_0, a_0, \tau$ ; Random:  $\text{diag}(TV_0, a_0)$

	Value	Std. Error	DoF	t-value	P-value
$TV_0$	220	5.4	3159	41	0
$a_0$	0.026	0.0020	3159	13	0
$\tau$	280	8600	3159	0.032	0.97

$$\frac{dV}{dt} = a_0 V, t \leq \tau; \frac{dV}{dt} = a_1, t > \tau. V \text{ is smooth: } \tau = \frac{1}{a_0} \log\left(\frac{a_1}{a_0 V_0}\right)$$

Fitting by NLME in R :  
 $TV_0 = 220 \text{ mm}^3$   
 $a = 0.026$   
 $\tau$ : Not sure

This fitting is based on all tumours. Looking at each histology may lead to different results.



# Logistic model fits well using SAEMIX

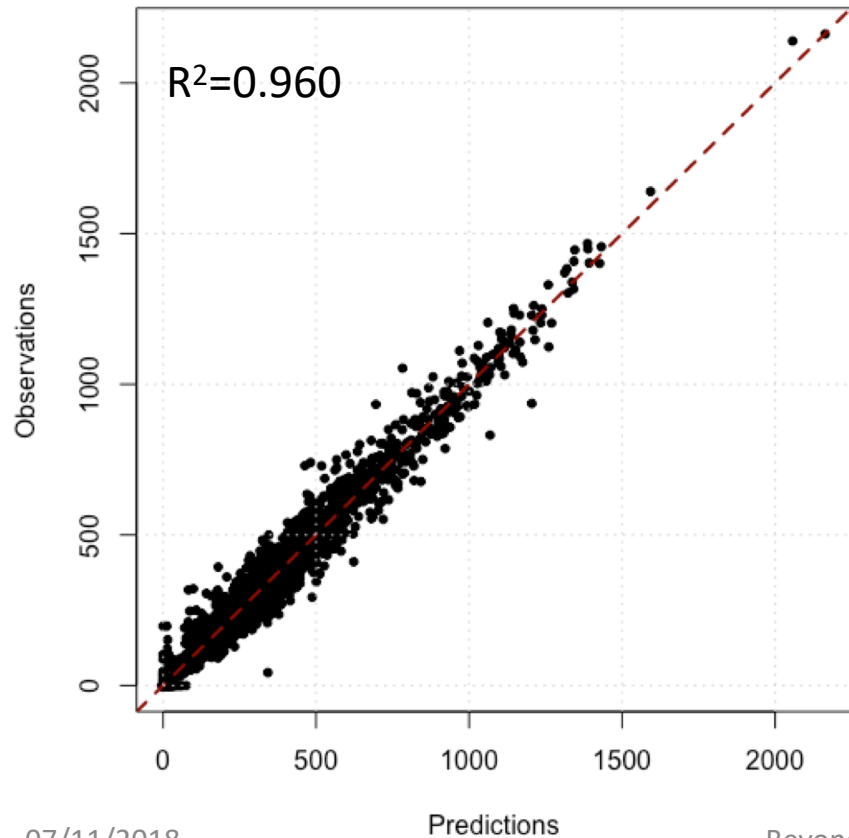
- Fixed:  $TV_0$ ,  $K$ ,  $a$ ; Random:  $\text{diag}(TV_0, K, a)$

$$\frac{dV}{dt} = aV \left( 1 - \frac{V}{K} \right)$$

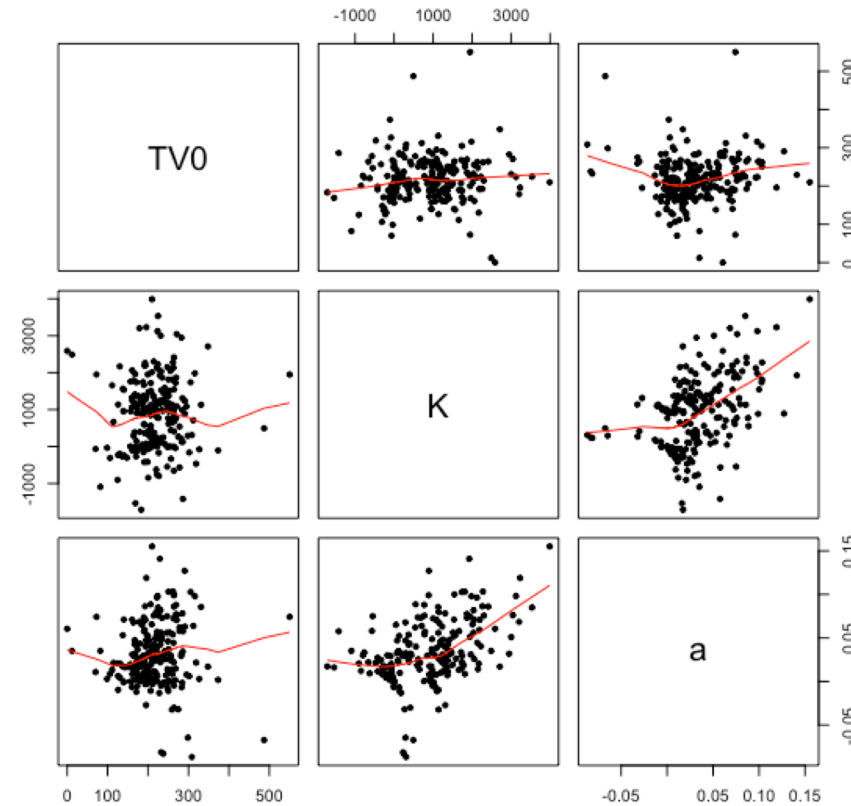
$a$ : growth rate (/day);  $K$ : carrying capacity ( $\text{mm}^3$ )

	Value	Std. Error	CV(%)
$TV_0$	220	4.7	2.2
$K$	950	160	16
$a$	0.032	0.0029	9.0

Fitting looks OK. Random effects not correlated  
 Logistic model can recapitulate all PDX's



Correlations between random effects



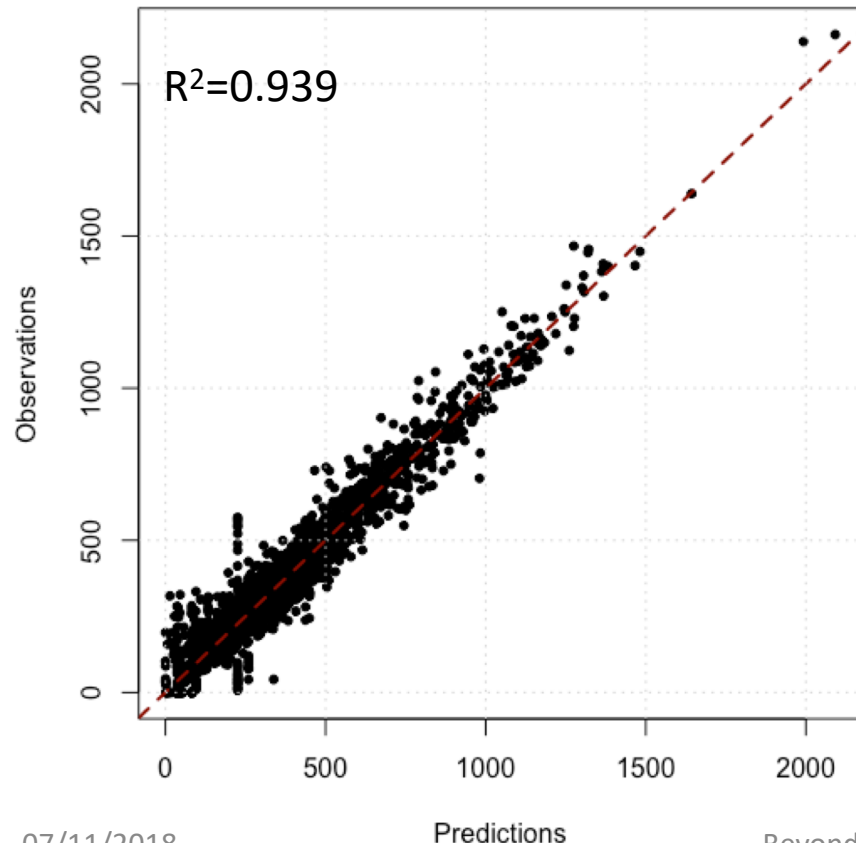
# Gompertz model was actually just exponential

- Fixed:  $TV_0, \alpha, \beta$ ; Random:  $\text{diag}(TV_0 + \alpha + \beta)$

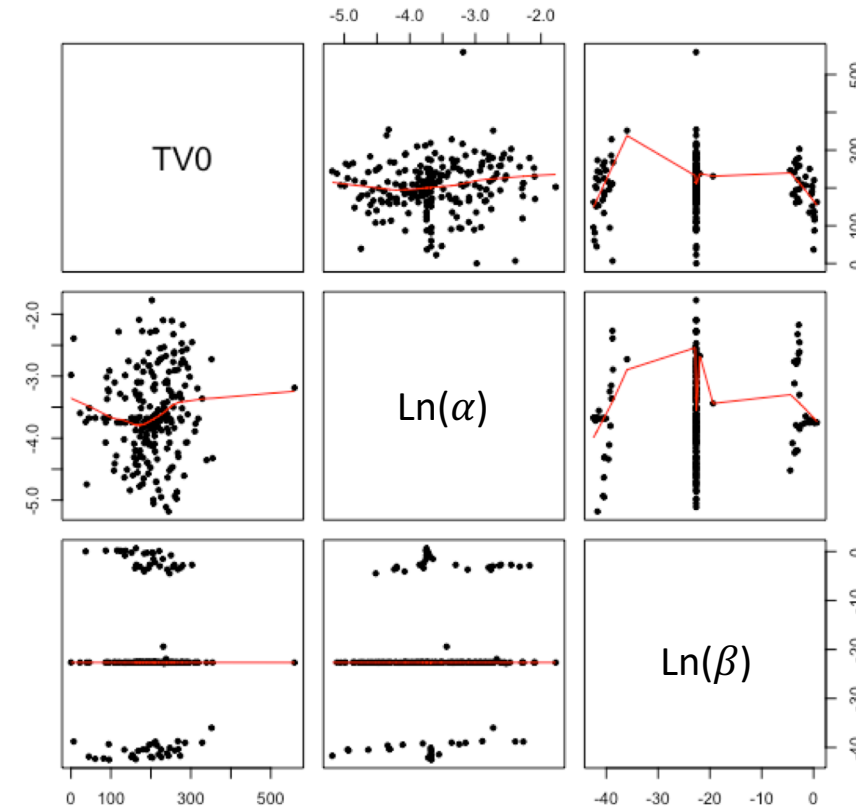
$$\frac{dV}{dt} = \alpha e^{-\beta t} V \quad \alpha: \text{growth rate (/day)}; \beta: \text{decay rate (/day)}$$

	Value	Std. Error	CV(%)
$TV_0$	210	5.0	2.4
$\text{Ln}(\alpha)$	-3.7	0.059	1.6
$\text{Ln}(\beta)$	-22.7	2.8	12

Fitting by SAEMIX:  $\beta$  is close to zero. Random effects not correlated. Gompertz reduced to exponential.

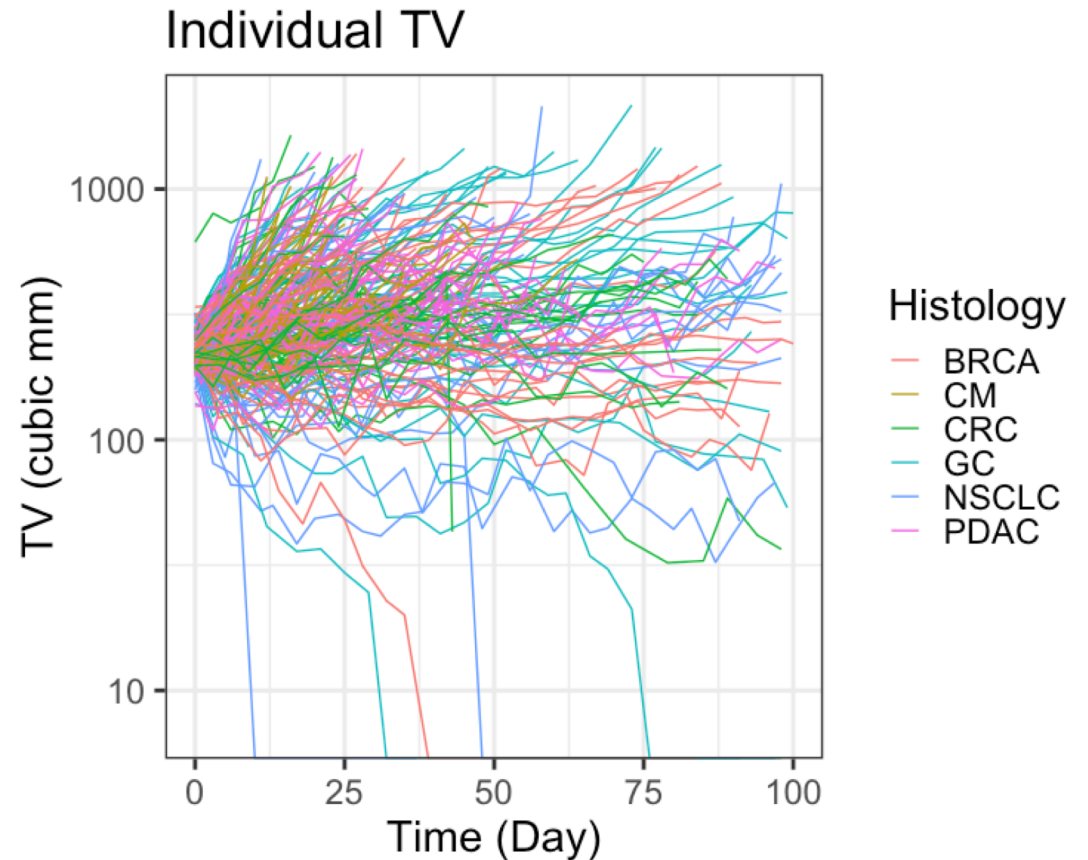
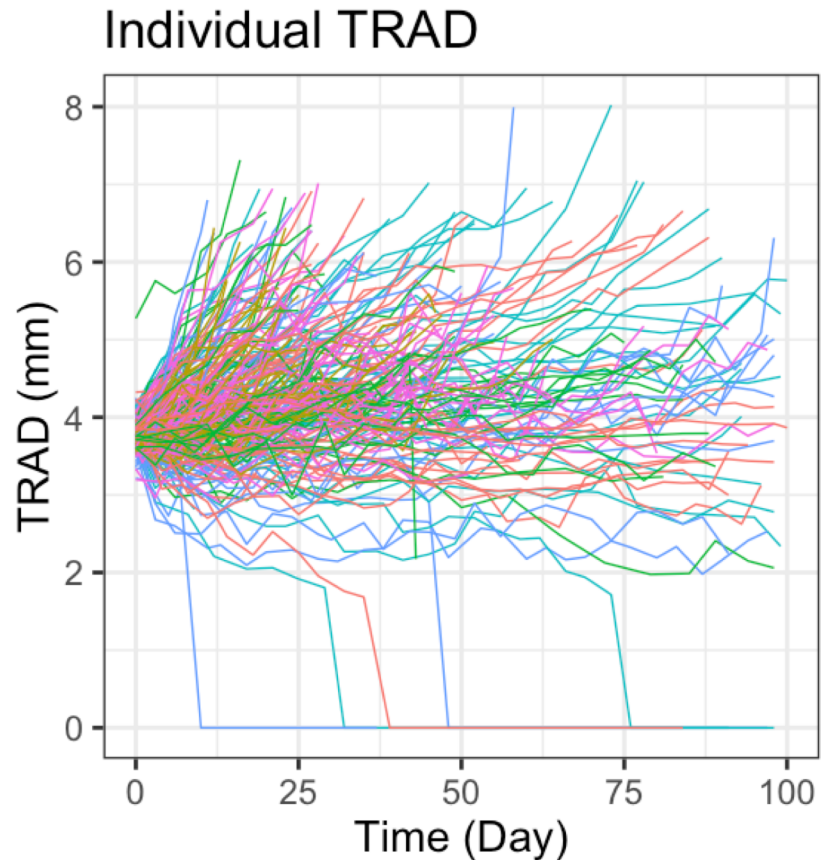


Correlations between random effects



# All rate laws accurately recapitulate control PDX growth

- Parametric inferences were successful
- Exponential-linear, logistic and Gompertz reduced to exponential model
- Essentially only two types of dynamics: linear and exponential



# All rate laws accurately recapitulate control PDX growth

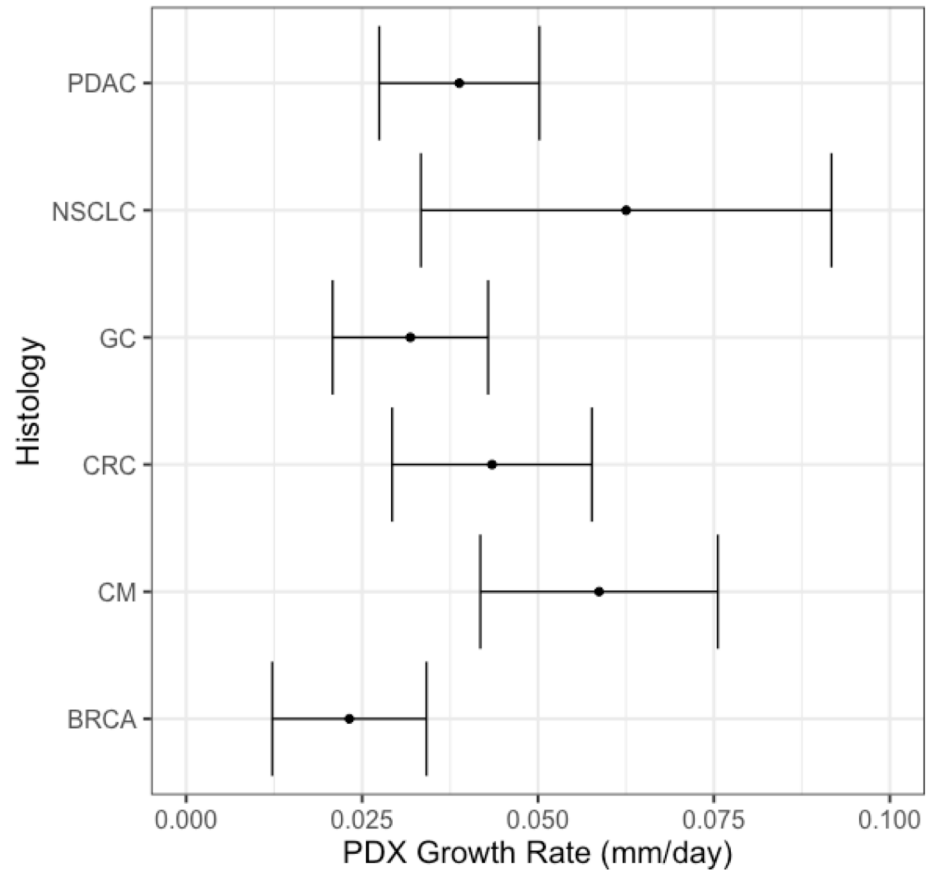
- Parametric inferences were successful
- Exponential-linear, logistic and Gompertz reduced to exponential model
- Essentially only two types of dynamics: linear and exponential
- Does growth rate vary by histology?
  - Inference using linear and exponential models
  - Are inference results consistent?



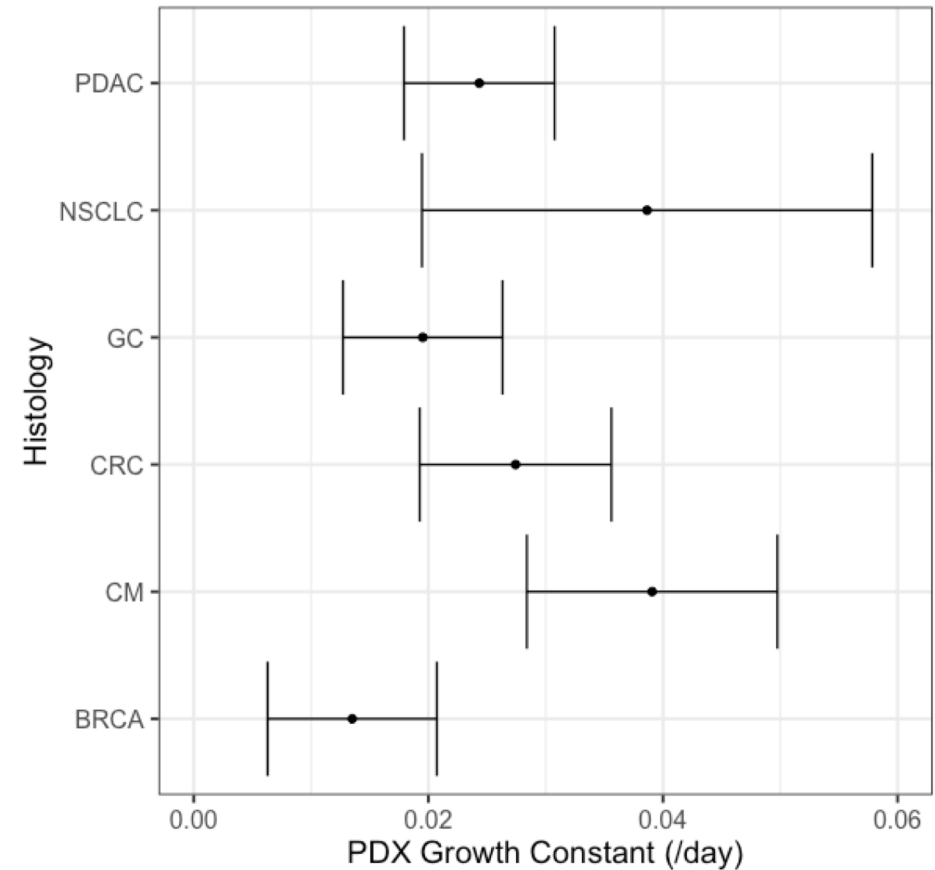
# Growth rate varies by histology

- Inferences by both models are largely consistent
  - Linear: NSCLC and CM grow significantly faster than BRCA
  - Exponential: PDAC, NSCLC, CM grow significantly faster than BRCA

PDX growth by histology (linear model)



PDX growth by histology (exponential model)



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